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NONLINEAR EFFECTS OF NANOPARTICLES: BIOLOGICAL VARIABILITY FROM HORMETIC DOSES, SMALL PARTICLE SIZES, AND DYNAMIC ADAPTIVE INTERACTIONS

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□ Researchers are increasingly focused on the nanoscale level of organization where biological processes take place in living systems. Nanoparticles (NPs, e.g., 1–100 nm diameter) are small forms of natural or manufactured source material whose properties differ markedly from those of the respective bulk forms of the “same” material. Certain NPs have diagnostic and therapeutic uses; some NPs exhibit low-dose toxicity; other NPs show ability to stimulate low-dose adaptive responses (hormesis). Beyond dose, size, shape, and surface charge variations of NPs evoke nonlinear responses in complex adaptive systems. NPs acquire unique size-dependent biological, chemical, thermal, optical, electromagnetic, and atom-like quantum properties. Nanoparticles exhibit high surface adsorptive capacity for other substances, enhanced bioavailability, and ability to cross otherwise impermeable cell membranes including the blood-brain barrier. With super-potent effects, nanoparticles can evoke cellular stress responses or therapeutic effects not only at lower doses than their bulk forms, but also for longer periods of time. Interactions of initial effects and compensatory systemic responses can alter the impact of NPs over time. Taken together, the data suggest the need to downshift the dose-response curve of NPs from that for bulk forms in order to identify the necessarily decreased no-observed-adverse-effect-level and hormetic dose range for nanoparticles.

Key words: nanoparticle, hormesis, nanomedicine, nonlinear dynamics, complex adaptive systems

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INTRODUCTION

Nanoparticles (NPs) are very small particles of material that may be natural or manufactured in origin (Buzea *et al.*, 2007; Ju-Nam and Lead, 2008; Merisko-Liversidge and Liversidge, 2011; Roduner, 2006). Sizes typically range from a fraction of one nanometer (nm) in diameter on at least one side up to 100 nanometers (European Commission on the Environment, 2011; International Organization for Standardization, 2005). Although submicron particles between 100–1000 nanometers in size have some advantages toward improving drug delivery (Oyewumi *et*

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al., 2010; Stovbun *et al.*, 2012; Wong, 2011), much of the focus of research interest in NPs has been on particles whose size falls below 100 nm. Their small size leads to a large surface area to volume ratio, resulting in variations of properties that differ markedly from those of bulk forms of the “same” material (Buzea *et al.*, 2007; Cao and Wang, 2011; Roduner, 2006).

The range of these altered effects encompasses electromagnetic, thermal, optical, biochemical, and even quantum (in quantum dots, at extremely small particle sizes, typically ranging from 1-10 nm, perhaps as high as 30 nm) properties. Smaller NPs readily cross cell membranes and translocate around the body via blood and lymph (Buzea *et al.*, 2007). Some NPs such as nano-silica or nano-silicon (Demento *et al.*, 2009; Mahony *et al.*, 2013; Petkar *et al.*, 2011; Wang *et al.*, 2012b) and nano-lipid carriers can also serve as immune adjuvants and markedly lower the amount of antigen needed to mount a response in the immune system, e.g., in one study to a dose as low as 2.5 nanograms (Bershteyn *et al.*, 2012; Diwan *et al.*, 2004).

In effect, nanoparticles are often biologically super-potent forms of their source material. For instance, the NP form of an antiretroviral drug in the 3 nanomolar range produced up to a 50-fold reduction in the 50% inhibitory concentration needed, compared with free drug doses (Chaowanachan *et al.*, 2013). Intermittent treatment with the nano-form of the immunosuppressant drug mycophenolic acid improves murine allograft survival time at a dose 1000-fold lower than the bulk form conventional drug (Shirali *et al.*, 2011). By extension, the therapeutic hormetic nanoparticle dose of an otherwise highly toxic source material might fall below the nanomolar level (Raja *et al.*, 2013), e.g., down to picomolar or even lower levels. Some studies further indicate the possibility of sinusoidal hormetic dose-response curves in such a situation (Malarczyk *et al.*, 2011).

Contemporary nanotechnology can generate manufactured nanoparticles in either a top down (e.g., milling, grinding) or bottom up (e.g., nano-silica self-assembly on a structural template) manner (Cho *et al.*, 2011; Cumbo *et al.*, 2013; Ju-Nam and Lead, 2008; Kiel *et al.*, 2012; Merisko-Liversidge and Liversidge, 2011). Reagents and manufacturing parameters such as solvents, dopants, coatings, biosynthetic plant extracts, pH, temperature, and sonication duration and intensity can influence the biological, chemical, electromagnetic, optical and physical properties of the resultant NPs (Cao and Wang, 2011; Pandey *et al.*, 2013; Roduner, 2006). Various nanoparticles will aggregate in the absence of capping agents and/or mechanical dispersion methods (Mudunkotuwa and Grassian, 2011; Pandey *et al.*, 2013; Pham *et al.*, 2007; Tang *et al.*, 2011; Zhang *et al.*, 2012).

APPLICATIONS IN MEDICINE AND TOXICOLOGY

In medicine, NPs have a growing importance for pharmacodiagnos-
tics and therapy (Ahn RW, 2013; Armstead and Li, 2011; Ho and Leong,
2010; Stark, 2011; Yoo *et al.*, 2011) as well as toxicology (Buzea *et al.*, 2007;
Winnik and Maysinger, 2013). Exemplar medical applications of nano-
forms are the enhanced drug or natural product delivery vehicles with
increased bioavailability and cell targeting potential in infectious diseases
(Armstead and Li, 2011; Dar *et al.*, 2013) and cancers (Al-Sadoon *et al.*,
2012; Chu *et al.*, 2012; Ghosh *et al.*, 2012; Sayed *et al.*, 2012; Shi *et al.*,
2010b; Wang and Thanou, 2010).

In the emerging area of theranostics, specialized nano-drugs enable
more precise targeting of specific cells and/or organs (Vivero-Escoto *et al.*,
2010). For instance, magnetic nanoparticle vehicles can be activated
to release their active agent for imaging and/or treatment only after they
enter their intended specific cancer cell target (Cole *et al.*, 2011; Ho *et al.*,
2011). The latter approach can take advantage of the nonlinear magnet-
ic behavior of the NPs (Geinguenaud *et al.*, 2012). Nonlinearity of
response can derive in part from the unique magnetic or optical proper-
ties of certain NPs. For example, near infrared light-activated cell-target-
ed nanoparticles can augment photothermal ablation therapies
(Melancon *et al.*, 2011). However, continuous wave versus nanosecond
pulsed laser stimuli can interact with nonlinear absorption processes of
gold nanospheres from plasmonic field enhancement to produce differ-
ent cell death mechanisms in the cancer cell nucleus versus cytoplasm
(Huang *et al.*, 2010).

Overall, NPs can more readily enter cancer cells because of the
increased vascular leakiness of tumors resulting in passive and/or active
uptake processes from ligands on the NP surfaces (Ghosh *et al.*, 2012; Sur
et al., 2010). Advantages of nano drug delivery vehicles with targeting
include a significant reductions of systemic toxicity in addition to lower-
ing total doses by orders of magnitude (Ahmad *et al.*, 2006; Armstead and
Li, 2011; Prakash *et al.*, 2010; Shirali *et al.*, 2011).

In environmental toxicology, high doses of many, though not all,
nanoparticles appear to be toxic and potentially contributory to a wide
range of diseases, from asthma to autoimmune diseases or atherosclerosis
and cancer (Buzea *et al.*, 2007; Winnik and Maysinger, 2013). Even
extremely low concentrations of nanoparticles of a given substance can
still exert toxic effects on model organisms e.g., 1 nanomolar ceria NPs on
C. elegans (Zhang *et al.*, 2011), sublethal adverse effects from 0.02 to 0.20
nanomolar silver NPs or 0.025 to 1.2 nanomolar gold NPs on developing
zebrafish embryos (Browning *et al.*, 2009; Lee *et al.*, 2012b; Osborne *et al.*,
2012; Truong *et al.*, 2013), or 20 nanograms/L silver NPs on juvenile
salmon (Farmen *et al.*, 2012). Across studies, silver NPs are overall more
toxic than gold NPs, but many particle- and organism-related variables

affect the specific findings. NP toxicity can derive from activating oxidative stress, inflammatory, immune and even apoptotic pathways as well as genotoxicity in cells (Gualtieri *et al.*, 2011; Sandberg *et al.*, 2012; Shi *et al.*, 2010a; Shi *et al.*, 2010b; Winnik and Maysinger, 2013). Plant-mediated biosynthesis of silver NPs can attenuate toxicity risk in some preparations (Barua *et al.*, 2013). One of the major questions that this heightened sensitivity from NPs asks is whether or not NPs violate the no-observed-adverse-effect level (NOAEL) principle of hormesis or is the NOAEL of NPs simply downshifted to lower levels and modified by the specific properties of a given NP form than with larger and bulk form particles?

HORMESIS AND NANOPARTICLES

At very low doses below the no-observed-adverse-effect level (NOAEL), previous investigators have documented evidence that some nanoparticles can initiate hormesis (Iavicoli *et al.*, 2010; Nascarella and Calabrese, 2012; Stovbun *et al.*, 2012). Hormesis is the nonlinear dose-response relationship in which low versus high doses of a given agent or stressor can exert effects in opposite directions. If an agent can inhibit function at a high dose, then the hormetic dose will stimulate function, and vice versa. Hormesis is increasingly understood as a dynamic adaptive response or biological plasticity of a complex living system at the level of the whole organism to intermittent mild stressors of various categories (Calabrese, 2013; Calabrese and Mattson, 2011; Iavicoli *et al.*, 2010). Types of stressor categories include physical, chemical, biological, and/or psychological factors (Calabrese and Mattson, 2011; Iavicoli *et al.*, 2010). Calabrese and Mattson (2011) have used hormesis as a quantitative estimate of biological plasticity.

For therapeutic applications, other researchers have proposed that exposing organisms to intermittently-timed hormetic stimuli could shift and shape epigenetic expression toward increased resilience against higher intensity stressors, disease, and aging itself (Stark, 2012; Vaiserman, 2010, 2011). Pickering *et al.* (2013) have emphasized the importance of spacing repeated exposures over time to permit expression of adaptive changes to oxidative stress. The beneficial effects of hormesis may arise from endogenous over-compensatory changes that the cell and organism use to repair or prepare for damage from larger magnitude, adaptively similar external threats from the environment (Stark, 2012; Van Wijk and Wiegant, 2010; Wiegant *et al.*, 2011).

The literature on cross-adaptation, cross-resistance and cross-sensitization includes many examples of overlapping phenomena that emphasize the role of biological plasticity mechanisms and time-dependent mechanisms in the organism. Pathways for the stress response networks involve interactions of metabolic, immune, inflammatory, hormonal, and autonomic functions. Thus, the biology of adaptation is an emergent

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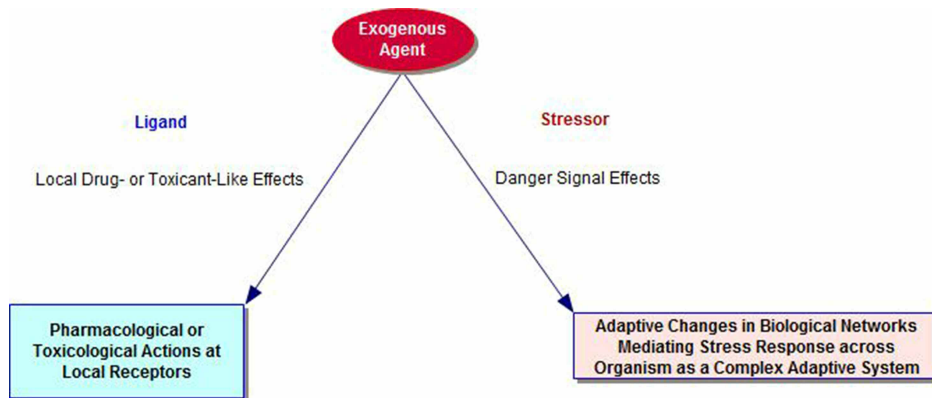


FIGURE 1. Dual Pathways for Exogenous Agents, including Nanoparticles, as Ligands and Stressors.

property of the organism as a whole and distinct from any local chemical effects of the exogenous agent on its specific receptors (see Figure 1). Sub-toxic and sub-lethal stressors initiate adaptive changes that can prepare the organism for future onslaughts from both the original stressor and agents where the physical nature, but not the elicited adaptive response repertoire of the recipient system, differs from that of the original (Milisav *et al.*, 2012). These latter types of responses are sometimes termed cross-adaptation (Hale, 1969) or cross-resistance (Milisav *et al.*, 2012) and, in other cases, heterologous hormesis (Calabrese *et al.*, 2007; Van Wijk and Wiegant, 2010).

However, there is a potentially important distinction for a specific agent and its dose or intensity level as a stressor for an individual organism. That is, some agents such as arsenic are relatively toxic and even lethal for most organisms at what pharmacologists consider low doses for their local effects. In that case, looking to a very low dose range might reveal a beneficial hormetic dose range for their adaptation-stimulating effects in most organisms. Thus, it makes sense to look for hormesis at extremely low doses of bulk form arsenic (Raja *et al.*, 2013) and perhaps even lower doses of nano-arsenic trioxide (Ahn *et al.*, 2013). Other nano-materials such as nano-calcium hydroxyapatite may be largely benign across a wider range of dose levels (Zhou and Lee, 2011) and/or can safely deliver nano-forms of less toxic agents for greater bioavailability and clinical benefit than reliance on more toxic bulk form drugs (Chun *et al.*, 2012; Joshi and Muller, 2009; Koning and Krijger, 2007; Lanao *et al.*, 2007; Moulari *et al.*, 2013; Zhao and Feng, 2010).

By analogy, for other types of more benign stressors such as exercise in physiology, the reaction to exercise as a “stressor” will also depend on the pre-established level of physical training and fitness of the individual. In general, exercise is not an inherently toxic event – rather, it is part of the physiological capacity of the organism. Nonetheless, an exercise level

that may be moderate for a highly fit person could even be lethal for a poorly-conditioned individual. Many doses of exercise may be fairly benign and initiate adaptive changes consistent with training effects. The “dose” of exercise that is beneficial will thus vary as a function of individual differences in the state of the organism at the time of the exercise. Genetic variations (Rodriguez *et al.*, 2012) and other environmental parameters (Lagisz *et al.*, 2013) can also influence individual differences in dose-response patterns for a variety of events and agents.

Certain NPs pose an additional challenge for determination of the no-observed-adverse-effect level (NOAEL) cut-off. In a complex nonlinear paradox, lower doses of NPs can sometimes increase rather than decrease the toxicity of a given source material as a function of the surface properties of the particles themselves. That is, in the absence of surface modifications to prevent spontaneous agglomeration from close physical interactions of highly reactive NPs in concentrated colloidal liquids (Bagwe *et al.*, 2006; Clark *et al.*, 2010; Sur *et al.*, 2012), higher concentrations or doses can favor NP agglomeration. The resultant nano-aggregates as a whole then can hide or quench the originally hyperreactive surfaces of their smaller NP “parts” (Mudunkotuwa and Grassian, 2011). Consequently, the specific larger agglomerated NP form is less toxic at a higher concentration than a lower dose of smaller, but well-dispersed NPs, e.g., NPs of PbS or copper.

For environmental toxicology, various forms of agitation, together with natural dilution factors that reduce concentration in the marine environment, for example, may disperse such agglomerates (Bourdineaud *et al.*, 2013; Rodrigues *et al.*, 2013; Ruan and Jacobi, 2012; Tang *et al.*, 2011; Zhang *et al.*, 2012). In such scenarios, the dispersion at a lower concentration re-exposes the hyperreactive surfaces of the smaller NPs and enhances their toxic potential (Mudunkotuwa and Grassian, 2011).

For nanomedicine applications, it is possible to take advantage of such issues by adding nontoxic capping agents (Singh *et al.*, 2013) and/or to time the use of sonication or vortexing prior to administration. Such a strategy can determine more systematically the nanoparticle size, shape, and surface chemistry. That is, certain capping agents, e.g., sugars or polysaccharides, can markedly reduce metal NP toxicity. Moreover, sonication will mechanically disperse any larger nanostructures that may have formed as a result of aging, agglomeration, and/or Ostwald ripening of the NPs in colloidal solution. In contrast, longer shelf storage over time could permit resumption of aging effects and thermodynamically-based development of increasingly larger nanostructures (Gautam *et al.*, 2013; Liu *et al.*, 2007). Thus, recency of sonication in solution can affect experimental and clinical findings (Bel Haaj *et al.*, 2013; Liu *et al.*, 2009; Murdock *et al.*, 2008; Tang *et al.*, 2011).

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Such issues become critical for evaluating NP effects in medicine and toxicology, when size, shape, and surface chemistry interact with dose to determine nonlinear response patterns (Mudunkotuwa and Grassian, 2011). For example, it is possible to reduce macrophage toxicity of anti-bacterial silver nanoparticles by coating the surfaces with chitosan (Jena *et al.*, 2012). Chitosan is a fibrous sugar extracted from shellfish outer skeletons. NP characteristics also contribute to unique challenges for controlling inter-experiment variability with NPs such as fullerene C60 nanoparticles (Chang and Vikesland, 2013).

What the data on nanomaterials raise for the discussion of hormesis is an analogous need to readjust our thinking about what constitutes a low dose, or more, precisely, a hormetic dose. Beyond the identity of the source material, the specific sizes, shapes, and surface charges of the NPs in air or water become significant factors interacting with dose. Hormesis researchers may need a type of sliding scale for defining very low hormetic doses where adverse events do not occur and yet beneficial adaptive changes can develop. First, the super-potent reactivity of small nanoparticles lowers the dose range for both toxic and, if relevant, therapeutic effects from a pharmacological perspective (Armstead and Li, 2011). Second, nanoparticles of the “same” material at a given low dose can exert either toxic or benign effects, depending in part on the size and surface reactivity of the particles (Lee *et al.*, 2012b; Mudunkotuwa and Grassian, 2011; Murdock *et al.*, 2008; Winnik and Maysinger, 2013).

Third, the coating or dopants on the surfaces of nanoparticles can also markedly change an otherwise toxic particle into a benign one or to acquire modified actions (Das *et al.*, 2013; McKibbin *et al.*, 2013; Rowe *et al.*, 2013; Sur *et al.*, 2012; Sur *et al.*, 2010; Thurber *et al.*, 2012; Tripathi *et al.*, 2009; Van Hoecke *et al.*, 2011). Fourth, the state of the recipient system cell or organism as a complex adaptive system (CAS) at the time of exposure is another modifying variable in the intensity and even direction of the response to nano and bulk form materials (Antelman and Caggiula, 1996; Bell and Schwartz, 2013; Browning *et al.*, 2009; Lee *et al.*, 2012b; Shi *et al.*, 2010b). Finally, NP forms of various source materials, including animal venoms, calcium phosphate, and nanocrystalline fullerene can exert marked toxicity for cancer cells but spare healthy cells (Al-Sadoon *et al.*, 2012; Harhaji *et al.*, 2007; Shi *et al.*, 2010b).

Thus, for nanoparticles, nonlinear hormetic responses are no longer a function of merely specific low doses. It is necessary to take into account the variable toxicity of a given specific nanoparticle based on its potential to change size, and hence, surface charge and related direct effects. The NPs then interact with individual differences in organisms’ ever-changing dynamical state of adaptive resilience to the lower dose ranges of nanoparticles in general. In general, the shift of the toxic dose range for nanoparticle forms of environmental toxicants toward lower doses is also

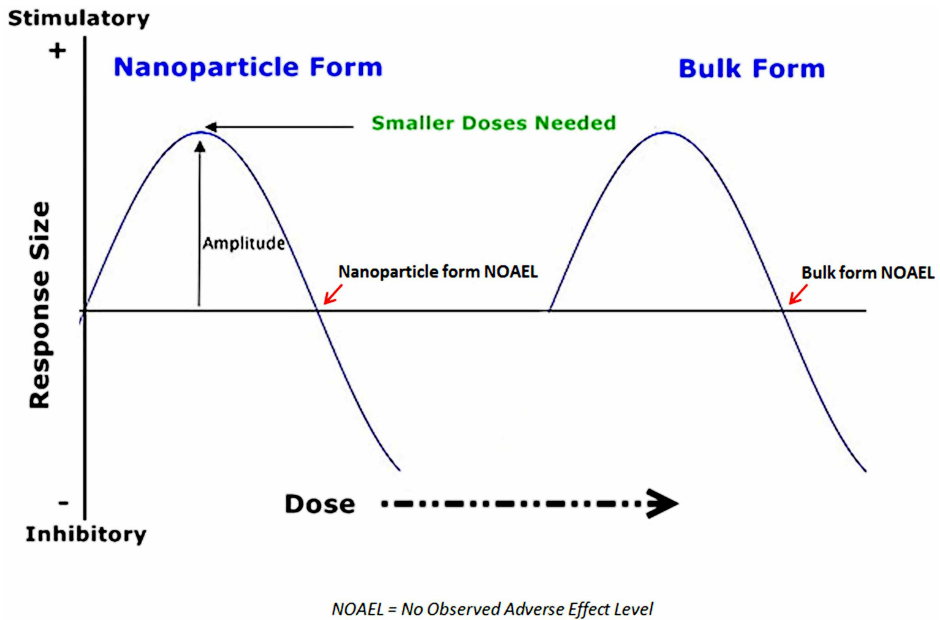


FIGURE 2. Left Shift of NOAEL and Hormetic Dose Range for Nanoparticles. Reprinted with permission (Bell and Schwartz, 2013).

orders of magnitude below that for bulk forms of the “same” materials. By extension, the no-observed-adverse-event-level (NOAEL) for nanoparticles of typically toxic agents must be very low to accommodate the range of various NP sizes and surface chemistries that may emerge within a given environmental context, far below the more fixed NOAEL for bulk forms of the same material (Figure 2). The potential for multiple particle-related, environment-related, and recipient-related factors to contribute variance in determining the NOAEL in each study of a given nanoparticle form make it much more difficult to define an appropriate metric for what constitutes a “low dose” or a hormetic dose.

Dose frequency also plays a role in adaptive phenomena. Nano-drugs persist inside cells longer than do conventional bulk form drugs (Ahmad and Khuller, 2008; Ahmad *et al.*, 2006; Armstead and Li, 2011; Shirali *et al.*, 2011). Even for bulk forms, with overly frequent dosing of an agent, the direction of the response can reverse when the nature of the response depends on the biological plasticity and metaplasticity mechanisms of adaptation to the agent as a stressor rather than on the mechanisms of its local effects on specific receptors (Abraham, 2008; Antelman and Caggiula, 1996; Antelman *et al.*, 1992; Antelman *et al.*, 2000; Pincus and Metten, 2010). Such data suggest the potential role of pulsed versus continuous dosing in mobilizing compensatory adaptive responses to an exogenous stressor or agent (Milisav *et al.*, 2012; Shirali *et al.*, 2011; Stark, 2012).

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In addition, the nature of adaptive responses, as opposed to local ligand-receptor responses, is that the emergent result is increased resistance to the original stressor and cross-resistance or cross-adaptation to other stressors that can affect similar adaptive pathways. In pharmacology and physiology research outside toxicology, many empirical examples of cross-adaptation, cross-resistance and cross-sensitization are documented (Antelman *et al.*, 1992; Antelman *et al.*, 2000; Hale, 1969; Milisav *et al.*, 2012). For instance, hypoxia cross-adapts with cold or hot temperatures (Banti *et al.*, 2008; Launay *et al.*, 2006; Lunt *et al.*, 2010; Ning and Chen, 2006); stress cross-sensitizes with amphetamine (Antelman *et al.*, 1980); sucrose cross-sensitizes with amphetamine or cocaine (Avena and Hoebel, 2003; Gosnell, 2005); formaldehyde cross-sensitizes with cocaine (Sorg *et al.*, 2001; Sorg *et al.*, 1998); heat shock, sodium arsenite, and cadmium chloride can cross-sensitize with one another depending on their heat shock protein activation patterns (Wiegant *et al.*, 1998).

The inference from such evidence is that an external agent at various doses is a salient biological stressor for the cell or organism as a complex adaptive system in addition to the local, receptor-specific actions (Bell and Schwartz, 2013). Given the data showing the ability of sub-toxic doses of NPs to initiate cellular stress responses, e.g., oxidative stress (Tang *et al.*, 2010; Winnik and Maysinger, 2013), cytokine and exosome release and other intercellular signaling events (Andersson-Willman *et al.*, 2012; Beloribi *et al.*, 2012; Demento *et al.*, 2009; Ristorcelli *et al.*, 2009; Zhu *et al.*, 2012a; Zhu *et al.*, 2012b), there are a number of potentially fruitful, albeit challenging directions for future research into biological mechanisms for NP-induced adaptive and hormetic responses (Demirovic and Rattan, 2013).

UNIQUE FEATURES OF NANOPARTICLES

Nonlinearity from Nanoparticle Properties: Beyond Hormetic Dose-Response Relationships

With nanoparticles, the nonlinearity of responses by cells and organisms may involve even more complexity than with bulk form materials. In contrast with conventional bulk forms of drugs, chemicals, herbs, and other materials, nanoscale forms vary in their effects as a function of not only the dose size, but also seemingly minor variations in their individual particle or aggregate sizes, shapes, and surface charges. Trace “contaminants,” “dopants,” and coatings on the surfaces of nanoparticles can also markedly change their properties, effects and level of toxicity at a given size (Isoda *et al.*, 2011; Kaur and Tikoo, 2012; Kumar *et al.*, 2012; Rowe *et al.*, 2013; Sun *et al.*, 2012; Sur *et al.*, 2012; Van Hoecke *et al.*, 2008; Wang *et al.*, 2012a). The environmental medium in which the NPs interact also modify their toxic or beneficial potential (Kaur and Tikoo, 2012; Mudunkotuwa and Grassian, 2011; Zhang *et al.*, 2012; Zhu *et al.*, 2006).

More highly charged surfaces often lead to greater NP toxicity (Truong *et al.*, 2013; Winnik and Maysinger, 2013). As a result, nanoparticles can evoke nonlinear response patterns from not only hormetic low dose-stimulatory response relationships, but also their inherent physico-chemical properties. That is, a given dose of NPs from the “same” source material can elicit lethal effects for one size nanoparticle, whereas the same dose at another size does not (Browning *et al.*, 2009; Lee *et al.*, 2012b).

In experimental cancer studies, certain sources of nanoparticles and certain sizes of those NPs are more toxic to cancer cells in vitro than to normal cells, e.g., calcium phosphate NPs (Shi *et al.*, 2010b). Enhanced intracellular access in “leaky” blood vessels supporting cancer cells, the inherently reactive surface properties of the NPs inside the cells, and different endogenous biological mechanisms may contribute to the differential cell type toxicity. Size-dependent responses and cell-specific interactions are a widespread phenomenon for NPs (Harhaji *et al.*, 2007; Jiang *et al.*, 2008; Kim *et al.*, 2012; Lankoff *et al.*, 2012).

In the real world environment, NP exposures encompass a wide range of particle sizes and shapes. Some NPs are crudely formed from uncontrolled environmental events that yield, irregular sizes, shapes and properties. Early laboratory methods for making nanoparticles involved prolonged grinding and milling procedures from bulk source materials (top-down methods), which make NPs with many structural irregularities and defects (DeCastro and Mitchell, 2002). Contemporary manufactured nanoparticles necessarily attain more consistent and defect-free shapes and sizes with more precise technological procedures like photo-lithography or bottom-up template synthesis methods (Ju-Nam and Lead, 2008).

In the manufacturing realm, nanotechnologists are also now using plants and other more natural biological agents to biosynthesize “green” silver and gold nanoparticles (Daisy and Saipriya, 2012; Das *et al.*, 2013; Hudecova *et al.*, 2012; Snitka *et al.*, 2012; Suriyakalaa *et al.*, 2012). Biologically synthesized silver and gold NPs have the advantage that the residual amounts of the plant extract adsorb onto the final NP surface and can enhance therapeutic effects while reducing toxicity (Tripathi *et al.*, 2009; Umashankari *et al.*, 2012). Some investigators include plant extracts, phytochemicals and antioxidants in their manufacturing methods to change surface properties and thereby reduce the cellular toxicity potential of certain metal NPs, e.g., silver NPs (Du *et al.*, 2012; Hudecova *et al.*, 2012; Lee *et al.*, 2012a; Mittal *et al.*, 2013; Osborne *et al.*, 2012; Park *et al.*, 2012; Suriyakalaa *et al.*, 2012; Tournebize *et al.*, 2012).

For more general sustainable NP manufacturing, natural plant materials such as English ivy, certain native plants from India, the traditional Chinese herb *Cuscuta chinensis*, glycyrrhizic acid from radix glycyrrhizae, guar gum, and rice husk also can also release their own organic nanoparticles of various sizes under appropriate conditions (Barve and

Chaughule, 2013; Burris *et al.*, 2012; Im *et al.*, 2011; Lenaghan *et al.*, 2013; Salavati-Niasari *et al.*, 2012; Soumya *et al.*, 2010; Wang *et al.*, 2013; Yen *et al.*, 2008). Beyond plant sources, a combination of ball-milling and ultrasound can reduce other organic material sources such as waste eggshells into calcium carbonate nanoparticles (Hassan *et al.*, 2013).

A rationale has been that all of the reagents used in making NPs will adsorb onto the large charged surface area to one degree or another and serve to “dope” or coat and thus modify the primary silver, gold, or silicon NP. Different solvents at different concentrations, for example, result in NPs of the “same” material with different characteristics and sizes (Abbasi and Morsali, 2012; Cao and Wang, 2011; Rao *et al.*, 2005; Yang *et al.*, 2011; Yoo *et al.*, 2006). As a result, the adsorbed materials can change the surface charge and properties of the NPs and thereby, their biological, therapeutic or toxic potential (Lu *et al.*, 2011; Sur *et al.*, 2010). The use of nontoxic or less toxic natural source reagents and materials could reduce toxic waste from NP manufacturing and potentially improve safety of nanomedicine products.

Studies on manufacturing also highlight the need for scrutiny of sample preparation and analytic methods in NP studies (Chikramane *et al.*, 2012). It is important to keep in mind that the precise laboratory conditions in which researchers examine effects of a nanoparticle preparation may influence the findings. In addition to any added solvents and reagents, basic manufacturing parameters such as intensity, duration, and the timing and extent of sonication will affect dispersion of nanoparticles that otherwise aggregate into larger particles with aging (Abbasi and Morsali, 2012; Murdock *et al.*, 2008; Ruan and Jacobi, 2012; Song *et al.*, 2012; Tang *et al.*, 2011). Variations in temperature and pH will also modify the properties of the nanoparticle samples in solution (Abbasi and Morsali, 2012; Rao *et al.*, 2005). Interactions with serum albumin lead to differential agglomeration and sizing of nanostructures during biological experiments (Song *et al.*, 2012; Tantra *et al.*, 2010).

Even making reliable NP concentrations for research purposes is potentially confounded by variations in additional nanostructures that might get into solution from sonication or vortexing agitation of the materials in liquid within different glassware or polymer containers (Betts *et al.*, 2013; Ives *et al.*, 2010; Liu *et al.*, 2012). Taken together, the data indicate the possibility of meaningful interactions between dose size, particle size, sample preparation and testing parameters, and state of the cells or organism at the time of administration in the expression of specific effects.

Quantum Properties of Nanoparticles

Smaller NP sizes also introduce quantum mechanical considerations into the problem for trying to evaluate nonlinear dose-response relationships in a conventional cause-effect medical model (Berec, 2012; Gupta

and Wiggers, 2011; Roduner, 2006; Yao and Hughes, 2009). For example, in nano-optics and nano-electronics, the smallest NPs (quantum dots) exhibit the ability to manifest macro quantum entanglement (Berec, 2012; Yao and Hughes, 2009), quantum coherence (Chudnovsky and Friedman, 2000; Hatef *et al.*, 2012), and quantum confinement (Gupta and Wiggers, 2011; Hannah *et al.*, 2012; Kleps *et al.*, 2010) phenomena. Some nanotechnologists take advantage of quantum confinement, for instance, to generate tunable quantum dot NPs with specific optical properties (Biju *et al.*, 2010; Kang *et al.*, 2011; Luther *et al.*, 2011; Troia *et al.*, 2009).

Few medical researchers consider the implications of quantum effects of NPs on biological function (Stovbun *et al.*, 2012). However, some investigators are using the ability of quantum confinement of electrons by different sizes of small NPs (quantum dots, with their atom-like properties) to yield different wavelength colors inside cells for specialized diagnostic imaging methods (Browning *et al.*, 2009; Huang *et al.*, 2012; Lee *et al.*, 2012b; McGuinness *et al.*, 2011; Shalchian *et al.*, 2005; Wang and Chen, 2011). Scientists working at the nanoscale point out the fact that biological processes occur at the nanoscale and sub-nanoscale level. Lloyd recently noted, for instance: “Nature is the great nano-technologist. The chemical machinery that powers biological systems consists of complicated molecules structured at the nanoscale and sub-nanoscale. At these small scales, the dynamics of the chemical machinery is governed by the laws of quantum mechanics” (Lloyd, 2011).

The literature contains only a few papers on the role of quantum physics in biological systems (Davies, 2004). Given limited evidence of the quantum mechanical properties of nanoparticles and their possible role in biological effects of NPs, much research lies ahead to understand the full potential effects of nanoparticles on living organisms. Nonetheless, at least some smaller sized NPs may act within the worlds of both conventional physics and quantum mechanical phenomena, making characterization of their dose-response relationships and mechanisms even more difficult to determine in a reproducible manner.

Still, it is important to realize that the biological effects of nanoparticles may change the scientific rules by which medical studies of nanoparticles are done. Clinical research on NPs could differ in design and even reliability from those for bulk forms of materials. The atom-like properties of very small nanoparticles may force quantum physics into the discussion of their therapeutic and toxicological effects in biology and medicine.

Interactions of NPs with the Organism as a Complex Adaptive System or Network

In the laboratory, technological advances now permit following the random walk of single nanoparticles through an individual organism, revealing the limitations to using averaged or ensemble data for evaluat-

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ing specific nanoparticle effects (Browning *et al.*, 2009; Lee *et al.*, 2012b). The latter issue of individual variability in NPs and individual differences in responses of each organism may hamper efforts to rely on the proposed *average* magnitude and response distributions for assessing hormesis from nanoparticles (Nascarella and Calabrese, 2012). Awareness of these factors, however, can reduce the risk of overly broad assumptions or generalizations about the effects of NPs in living systems, including relative to hormesis.

So far, this paper has alluded to the complex nonlinear dynamics and network organization of the organism as another factor in modifying the nature, magnitude and direction of nanoparticle effects (Sugarman *et al.*, 2013). What are the implications of considering the interaction of the nanostructured material with an individual organism? In his text, *Introduction to Nanoscience*, Lindsay (Lindsay, 2010) commented: “Nanoscience is where atomic physics converges with the physics and chemistry of complex systems.”

Current thinking suggests that the adaptive response nature of hormesis is a manifestation of biological plasticity (Calabrese and Mattson, 2011). As a result, the dose-response observations reflect emergent interactions of a given mild stressor or low dose agent with a specific organism in a particular dynamic state, modified by genetics and past experiences. Biological metaplasticity, or the plasticity of plasticity can reverse directionality of responses as a function of past adaptations around a set point compatible with maintaining homeostatic balance (Abraham, 2008; Antelman and Caggiula, 1996). Moreover, for *in vivo* studies, NPs may yield very different findings from *in vitro* experiments (Clift *et al.*, 2011; Lu *et al.*, 2011). Using intact organisms may ultimately be necessary to understand when and how NPs might cause hormesis and other nonlinear responses, e.g., stochastic resonance (Chen *et al.*, 2012; McDonnell and Abbott, 2009).

Complete organisms and intact cells are each complex adaptive systems (CAS) at different levels of scale. CASs are interconnected, interactive and interdependent networks of self-organized components. The specialized components in turn generate emergent properties at the higher levels of organization not seen in the individual parts (Pincus and Metten, 2010). Furthermore, a CAS can change behaviors over time at different time scales, with a range between order and chaos that adapts nonlinearly to changes in the environment.

The result observed can vary, revealing degrees of resilience from the capacity for adjusting intrinsic flexibility and stability in the behaviors of the complex system. The dynamic self-organized “goal” for a CAS is to optimize the organism or cell’s fitness within a given environment to the extent possible within the current state and meta-flexibility of the system (Pincus and Metten, 2010). The nature, magnitude, and direction of the

change can be difficult to predict in a CAS, especially at critical points of dynamical instability (Hollenstein, 2007; Malarczyk *et al.*, 2011; Sugarman *et al.*, 2013). In addition, Figure 3 illustrates some time-dependent variables in a living CAS, e.g., developmental state of the recipient organism and the frequency of repeated exposures that can influence the adaptive changes and even the direction of the observed responses, apart from the dose itself.

Some researchers in nonlinear dynamical systems (NDS) propose that disease and aging reflect losses of complexity in the dynamics of the organism (Costa, 2002, 2007; Costa *et al.*, 2002; Fredrickson and Losada, 2005; Goldberger *et al.*, 2002; Losada, 1999; Losada and Heaphy, 2004).

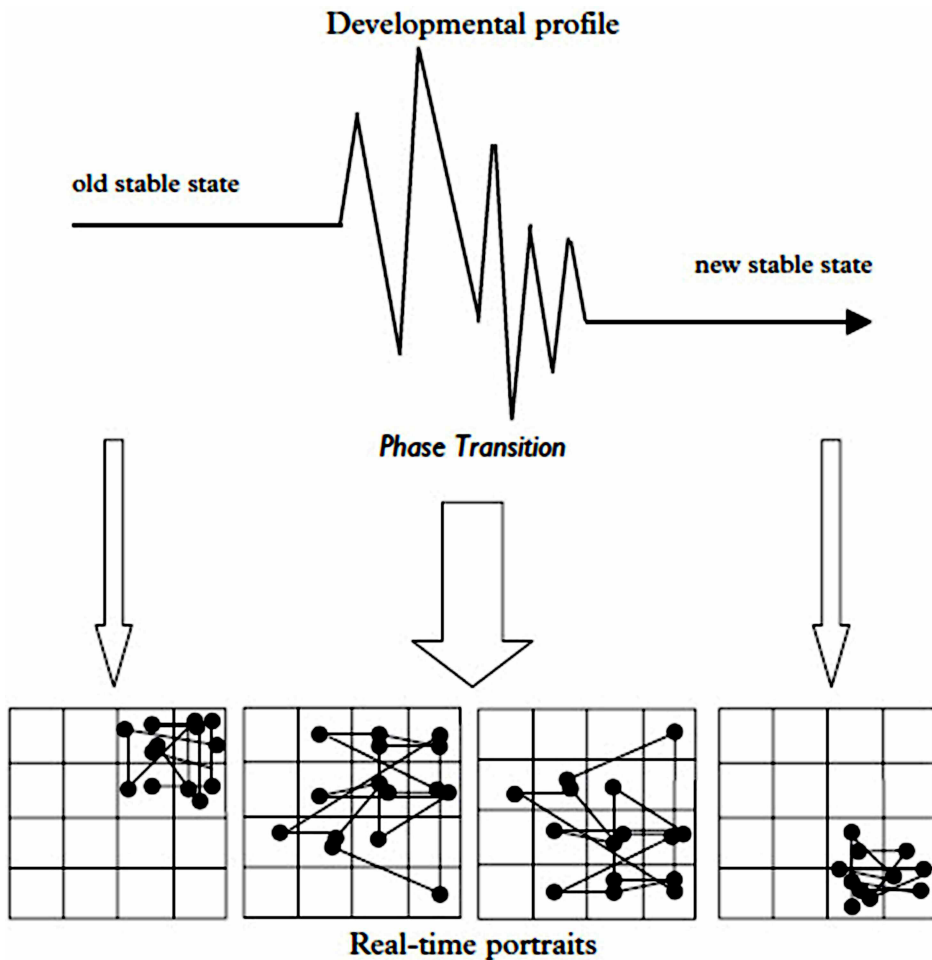


FIGURE 3A. Time-Dependent Interactions of Host and Environment that Can Affect the Initiation and Direction of Responses in a Nonlinear Dynamical System such as a Living Organism. Developmental phase transition in an adolescent human being can destabilize system dynamics and lead to subsequent self-reorganization of interpersonal interaction dynamics over time. Reprinted with permission (Hollenstein, 2007).

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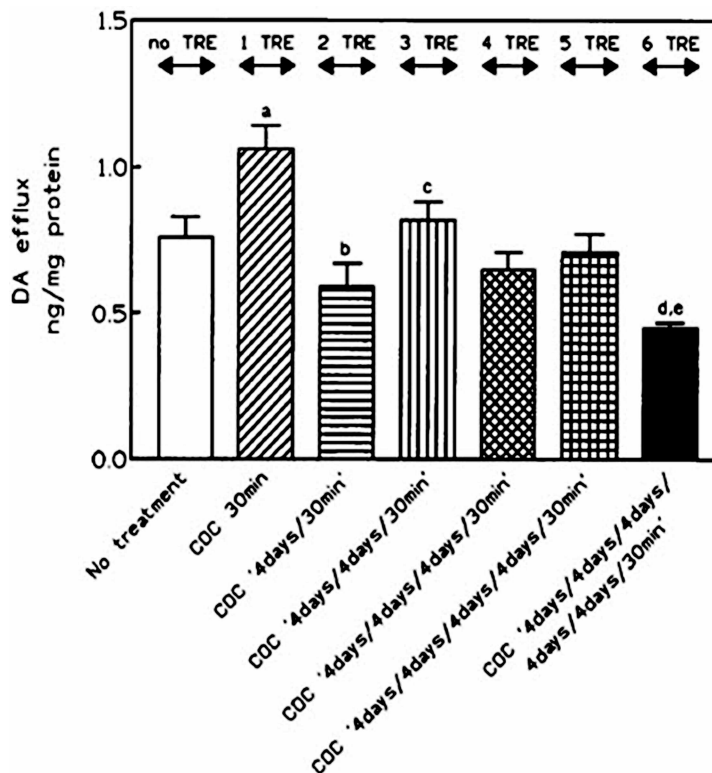


FIGURE 3B. Overly frequent repetitions of a low dose environmental stressor (e.g., experimental cocaine pretreatments in an animal during a sensitization protocol study of brain striatal dopamine efflux evoked by eliciting dose of cross-sensitized amphetamine) leads to reversal of direction in observed responses over consecutive repetitive exposures. Reprinted with permission (Antelman et al., 1997).

With regard to high doses of nanoparticles and other small particles from air pollution, many believe that such toxic level exposures promote disease, as reflected in a loss of complexity in physiological measures such as heart rate variability (Shannahan *et al.*, 2012).

However, studies on the effects of lower, subtoxic NP doses reveal individual variability in the effects of the agent on different organisms receiving the “same” exposure. Stressing a CAS and observing how it responds to the stressor in spatially and temporally remote areas of function can reveal the larger capacity for resilience of the individual organism (Bar-Yam, 1997; Bar-Yam and Epstein, 2004). Cause-effect relationships in CAS tend to be indirect rather than direct. Overwhelming intensities of stress can completely disrupt a complex biochemical network, for instance, whereas lesser levels of stress may simply induce a self-reorganization of function to cope with the effects (Mihalik and Csermely, 2011; Szalay *et al.*, 2007) (Figure 4).

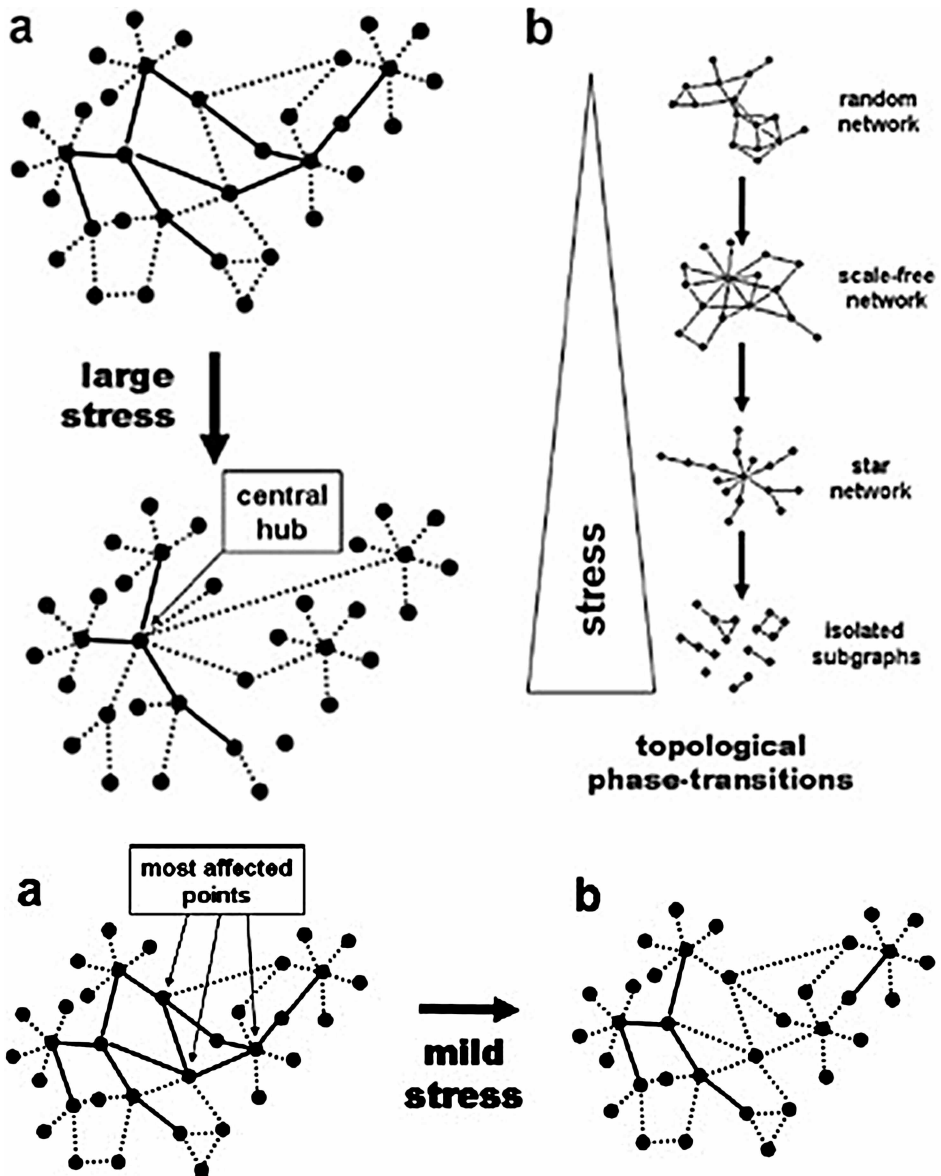


FIGURE 4. Effects of Different Levels of Stress on Functional Network Organization of a Complex Adaptive System. Reprinted with permission (Szalay *et al.*, 2007). **Note:** Solid and dotted lines represent strong and weak (high and low affinity) links, respectively.

For instance, with silver NP exposures at sub-lethal doses, some developing zebrafish organisms remain healthy whereas others develop deformities (Lee *et al.*, 2012b). Those with the deformities show an NP size-dependent effect, with larger numbers of larger versus smaller size silver NPs accumulating inside the surviving deformed versus healthy individuals, at the same given molar concentration. Such data support the likely

interactions of subtoxic nanoparticle doses and sizes with the state of the organisms at the time of exposure. The dynamical state of the individual CAS here would translate into the variable ability of blood vessel integrity and cell membranes either to allow or to block entry of the damaging larger-sized nanoparticles inside the cells, e.g., (Shi *et al.*, 2010b).

In the therapeutic realm, harnessing low doses of certain sized nanoparticles as hormetic stimuli may be useful. It is instructive to look at research on using low level discrete, well-timed stimuli to mobilize widespread changes in function of the overall organism. For example, adding a low level, subsensory noise applied to the feet of elderly individuals can improve the complexity of sway fluctuations in their postural balance (Costa, 2007). Adding noise in the latter case enhances the ability to detect otherwise age-weakened sensory signals within the organism. On the other hand, applying discrete pulsed electrical stimuli with precise magnitude and timing for the individual's diseased state (i.e., which is emergent biological "noise") can disrupt and normalize cardiac arrhythmias or experimental epileptic seizure activity in the brain (Coffey, 1998; Garfinkel *et al.*, 1992; Schiff *et al.*, 1994).

One underlying theory for the clinical benefits of introducing a small or weak signal within a larger endogenous noise in a CAS is stochastic resonance (SR) (Casado-Pascual *et al.*, 2003; Czaplicka *et al.*, 2013; Kelty-Stephen and Dixon, 2013; Korn and Faure, 2003; Krawiecki *et al.*, 2000; Magalhaes and Kohn, 2011; McDonnell and Abbott, 2009; Pinamonti *et al.*, 2012; Torres and Ruiz, 1996). SR is a phenomenon which involves the ability of a small signal to exert noise-enhanced amplified effects when given in the background of the much larger noise to a nonlinear complex system (Figure 5) (McDonnell and Abbott, 2009).

Certain nanoparticles, e.g., carbon nanotube transistors, can evoke this type of noise-amplified response to a weak signal in a non-living complex system (Lee *et al.*, 2006). In biological systems, previous studies have demonstrated stochastic resonance in sensory systems. SR is involved, for example, in crayfish detection of incident pressure waves from predators as well as in human visual perception and balance control (Moss *et al.*, 2004). To further explore SR-related phenomena, Lee *et al.* (2010) showed coherence resonance or self-synchronization at an optimal noise level, in transport of single ions through the interior of a 500 micrometer long carbon nanotube. The latter observations involved increases in throughput of the nanopore by a factor of 100 (Lee *et al.*, 2010).

SR is a testable hypothesis as one way in which a pulsed dose of a salient, low dose agent or nanoparticle might initiate the cascades of amplified biological signaling reported in hormesis (Calabrese, 2013). The "noise" in an adaptive living system might be a pattern of dysfunctions manifesting as a disease, toxicity, or aging (Soti and Csermely, 2007). Then the therapeutic strategy could be either (a) to add noise to enhance sen-

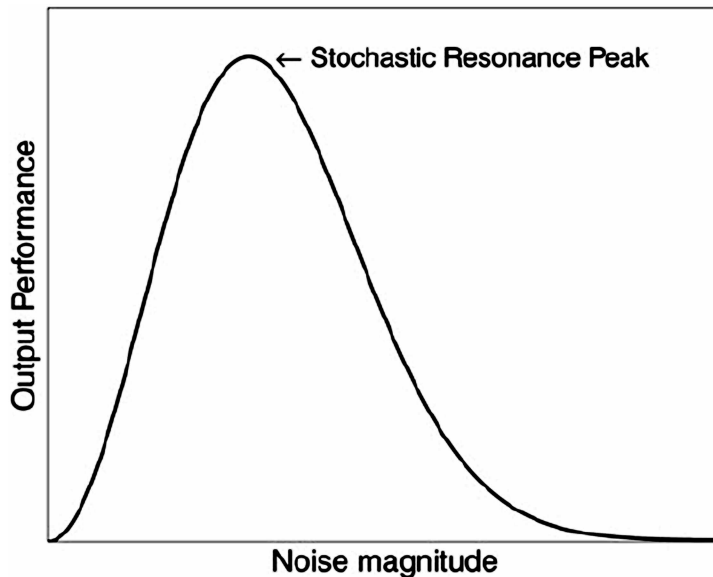


FIGURE 5. Nature of Stochastic Resonance (SR) as a Model Nonlinear Process for Noise-Enhanced Small Signals. Typical SR Curve of Output Performance versus Input Noise Magnitude for Complex Systems Capable of Stochastic Resonance. Reprinted under the Creative Commons License with Attribution (McDonnell and Abbott, 2009).

sory detection capacity in an aging individual (Costa *et al.*, 2007) or (b) to introduce a salient mild hormetic signal into the pre-existing systemic noise of disease to trigger a reversal of direction toward health (Stark, 2012; Torres and Ruiz, 1996; Van Wijk and Wiegant, 2011; Yu *et al.*, 2013).

SR may play a role in the aging process and in anti-aging interventions. Soti and Csermely (2007) have proposed that aging leads to increased noise in the functional cellular biochemical networks. The noise grows via cumulative damage to weak biochemical network links involving chaperone proteins such as heat shock proteins. Both aging and disease can induce a loss of complexity in the nonlinear dynamics of a complex adaptive system across levels of organizational scale, including cell systems, physiological systems, and whole organisms (Costa, 2007; Costa *et al.*, 2005; Fredrickson and Losada, 2005; Goldberger, 1996; Goldberger *et al.*, 2002; Hollenstein, 2007; Pincus and Metten, 2010; Soti and Csermely, 2003, 2007).

However, well-timed mild hormetic stressors from certain nanoparticles in low doses and certain particle sizes could serve as one type of small salient SR signal embedded in the larger noise to trigger beneficial recovery of complexity in the system (Stark, 2012; Sugarman *et al.*, 2013). Modulation of heat shock proteins offers a potential biological mechanism (Soti and Csermely, 2006) by which to reverse age- or disease-related loss of complexity in the adaptive networks of cells. In hormesis research, one intervention strategy involves postconditioning hormesis to

elicit therapeutic effects in heat shock protein activation patterns from mild (low dose) hormetic environmental stimuli (Van Wijk and Wiegant, 2010; Wiegant *et al.*, 2011). Previous studies have already shown the capacity of various nanoparticles at toxic doses to modulate heat shock protein activation patterns (Farmen *et al.*, 2012; Foldbjerg *et al.*, 2012; Lim *et al.*, 2012; Richert *et al.*, 2012; Siddiqi *et al.*, 2012; Zhao *et al.*, 2012).

It is not always necessary to use low doses of the same specific stressor that may have caused deterioration. In the adaptive stress response networks, the phenomenon of cross-adaptation or cross-resistance (Hale, 1969; Milisav *et al.*, 2012) could permit selection of a heterologous, cross-adapted stressor to serve as the hormetic stimulus. Newer evidence suggests that low doses of certain salient cross-adapted nanoparticles could act as such postconditioned hormetic stressors (Bell and Schwartz, 2013). It may also be possible, as Vaiserman has proposed, to use intermittent mild hormetic stressors to initiate preconditioned hormesis and adaptive changes for preventive purposes (Vaiserman, 2010, 2011). Nonetheless, identifying the optimal conditions for beneficial shaping of health- and longevity-promoting exposures remains a challenge (Pickering *et al.*, 2013).

For low doses of small nanoparticles to act via stochastic resonance, e.g., in hormesis, they would need to take advantage of endogenous amplification processes possible within the individual as a nonlinear complex adaptive system (McDonnell and Abbott, 2009). Notably, the NPs would need to arrive as a discrete properly-timed, pulsed low intensity signal rather than at continuous dosing levels (Antelman *et al.*, 2000; Casado-Pascual *et al.*, 2003; Kelty-Stephen and Dixon, 2013). While speculative, the concept of stochastic resonance in complex adaptive systems could add a new layer of discovery to advance our understanding of the circumstances in which the effects of hormesis might be utilized for prevention or treatment of disease.

CONCLUSIONS

In conclusion, for the therapeutic application of hormesis with NPs (Iavicoli *et al.*, 2010; Nascarella and Calabrese, 2012), additional considerations beyond traditional dose explanations likely come into play to understand nonlinear responses. Numerous interacting factors related to nanoparticle size, shape, and surface charge, in addition to the material composition and low dose, determine the net effects of particular nanoparticles in a given study. The NPs then interact with individual differences in the dynamical state of the cells and organisms as complex adaptive systems to generate emergent nonlinear effects.

The time-dependent, multifactorial and individualized nature of adaptive phenomena raises significant questions about the most appropriate experimental designs on NP hormesis. Careful characterization of nanoparticles used in a given study and the pre-treatment dynamical state

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of the individual recipient organisms or cells may lessen the risk of generating confusing and even irreproducible findings in this field (Chang and Vikesland, 2013; Vijayaraghavan and Nalini, 2010; Xia *et al.*, 2009). Even relying on averaged versus individualized data may be misleading (Browning *et al.*, 2009; Huang *et al.*, 2012; Lee *et al.*, 2012b).

Implications of the available literature for future studies on NPs and hormesis include:

- Because of enhanced biological potencies of NPs, it is necessary to look for significant down-shifting to the left along the x-axis for dose levels below the no-observed-adverse-effect-level (NOAEL) where hormesis is more likely to occur. That is, in some cases, hormetic doses of some NPs may sometimes occur at levels below 1 nanomolar concentration. However, since NP size and surface chemistries can vary in a given environment, the cut-off levels for their direct and indirect therapeutic and/or toxic effects may also vary accordingly.
- Because of the potential interactions of small particle size and low dose with the state of the recipient complex adaptive system, developing multifactorial models for biological plasticity mechanisms of hormesis may be particularly important with nanoscale materials.
- For therapeutic applications of hormesis using NPs, dosing regimens may need to involve discrete pulsed rather than continuous administration of the low doses of specific sized NPs in order to take advantage of biological signaling and nonlinear stochastic resonance. Timing of NPs can affect the nature and direction of the response (Hossu *et al.*, 2010; Jonasson *et al.*, 2013; Vesterdal *et al.*, 2010). The amplified effects of small pulsed signals in the context of a larger noisy signal can produce large magnitude, clinically significant change in an individual as a complex adaptive system (Costa, 2007; Ichiki and Tadokoro, 2013; McDonnell and Abbott, 2009; Pinamonti *et al.*, 2012; Soti and Csermely, 2007).

The past decade has seen an explosion of research and discovery in nanoscience, nanotechnology, and nanomedicine. The potential of interacting therapeutic nanoparticles with hormetic dose treatment strategies is largely unexplored. Tools from systems biology (Abu-Asab *et al.*, 2011), network analysis (Farkas *et al.*, 2011) and nonlinear dynamical systems (Pincus and Metten, 2010) may facilitate this direction for future research. The evidence supports the importance of exploratory and hypothesis-driven studies in this area.

REFERENCES

- Abbasi AR and Morsali A. 2012. Influence of solvents on the morphological properties of AgBr nanostructures prepared using ultrasound irradiation. *Ultrason Sonochem* 19:540-545
- Abraham WC. 2008. Metaplasticity: tuning synapses and networks for plasticity. *Nat Rev Neurosci* 9:387

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- Abu-Asab MS, Chaouchi M, Alesci S, Galli S, Laassri M, Cheema AK, Atouf F, VanMeter J, and Amri H. 2011. Biomarkers in the age of omics: time for a systems biology approach. *OMICS*. 2011 Mar;15(3):105-12. Epub 2011 Feb 14.
- Ahmad Z and Khuller GK. 2008. Alginate-based sustained release drug delivery systems for tuberculosis. *Expert Opin Drug Deliv* 5:1323-1334
- Ahmad Z, Pandey R, Sharma S, and Khuller GK. 2006. Alginate nanoparticles as antituberculosis drug carriers: formulation development, pharmacokinetics and therapeutic potential. *Indian J Chest Dis Allied Sci* 48:171-176
- Ahn RW, Barrett SL, Raja MR, Jozefik JK, and Spaho L. 2013. Nano-Encapsulation of Arsenic Trioxide Enhances Efficacy against Murine Lymphoma Model while Minimizing Its Impact on Ovarian Reserve In Vitro and In Vivo. *PLoS ONE* 8:e58491
- Al-Sadoon MK, Abdel-Maksoud MA, Rabah DM, and Badr G. 2012. Induction of Apoptosis and Growth Arrest in Human Breast Carcinoma Cells by a Snake (*Walterinnesia aegyptia*) Venom Combined With Silica Nanoparticles: Crosstalk Between Bcl2 and Caspase 3. *Cell Physiol Biochem* 30:653-665
- Andersson-Willman B, Gehrmann U, Cansu Z, Buerki-Thurnherr T, Krug HF, Gabrielsson S, and Scheynius A. 2012. Effects of subtoxic concentrations of TiO₂ and ZnO nanoparticles on human lymphocytes, dendritic cells and exosome production. *Toxicol Appl Pharmacol* 264:94-103
- Antelman SM and Caggiula AR. 1996. Oscillation follows drug sensitization: implications. *Critical Reviews in Neurobiology*. 10:101-117
- Antelman SM, Caggiula AR, Gershon S, Edwards DJ, Austin MC, Kiss S, and Kocan D. 1997. Stressor-induced oscillation. A possible model of the bidirectional symptoms in PTSD. *Annals of the New York Academy of Sciences*. 821:296-304
- Antelman SM, Caggiula AR, Knopf S, Kocan DJ, and Edwards DJ. 1992. Amphetamine or haloperidol 2 weeks earlier antagonized the plasma corticosterone response to amphetamine; evidence for the stressful/foreign nature of drugs. *Psychopharmacology*. 107:331-336
- Antelman SM, Eichler AJ, Black CA, and Kocan D. 1980. Interchangeability of stress and amphetamine in sensitization. *Science*. 207:329-331
- Antelman SM, Levine J, and Gershon S. 2000. Time-dependent sensitization: the odyssey of a scientific heresy from the laboratory to the door of the clinic. *Molecular Psychiatry*. 5:350-356
- Armstead AL and Li B. 2011. Nanomedicine as an emerging approach against intracellular pathogens. *Int J Nanomedicine* 6:3281-3293
- Avena NM and Hoebl BG. 2003. Amphetamine-sensitized rats show sugar-induced hyperactivity (cross-sensitization) and sugar hyperphagia. *Pharmacol Biochem Behav* 74:635-639
- Bagwe RP, Hilliard LR, and Tan W. 2006. Surface modification of silica nanoparticles to reduce aggregation and nonspecific binding. *Langmuir* 22:4357-4362
- Banti V, Loreti E, Novi G, Santaniello A, Alpi A, and Perata P. 2008. Heat acclimation and cross-tolerance against anoxia in *Arabidopsis*. *Plant Cell Environ* 31:1029-1037
- Bar-Yam Y. 1997. *Dynamics of Complex Systems*. Perseus Books, Reading, MA
- Bar-Yam Y and Epstein IR. 2004. Response of complex networks to stimuli. *Proceedings of the National Academy of Sciences of the United States of America* 101:4341-4345
- Barua S, Konwarh R, Bhattacharya SS, Das P, Devi KS, Maiti TK, Mandal M, and Karak N. 2013. Non-hazardous anticancerous and antibacterial colloidal 'green' silver nanoparticles. *Colloids Surf B Biointerfaces* 105:37-42
- Barve R and Chaugule R. 2013. Size-dependent in vivo/in vitro results of homeopathic herbal extracts. *Journal of Nanostructure in Chemistry* 3:18
- Bel Haaj S, Magnin A, Petrier C, and Boufi S. 2013. Starch nanoparticles formation via high power ultrasonication. *Carbohydrate polymers* 92:1625-1632
- Bell IR and Schwartz GE. 2013. Adaptive network nanomedicine: an integrated model for homeopathic medicine. *Frontiers in Bioscience (Scholar Ed.)* 5:685-708
- Beloribi S, Ristorcelli E, Breuzard G, Silvy F, Bertrand-Michel J, Beraud E, Verine A, and Lombardo D. 2012. Exosomal Lipids Impact Notch Signaling and Induce Death of Human Pancreatic Tumoral SOJ-6 Cells. *PLoS One* 7:e47480
- Berec V. 2012. Quantum entanglement and spin control in silicon nanocrystal. *PLoS One* 7:e45254
- Bershteyn A, Hanson MC, Crespo MP, Moon JJ, Li AV, Suh H, and Irvine DJ. 2012. Robust IgG responses to nanograms of antigen using a biomimetic lipid-coated particle vaccine. *J Control Release* 157:354-365

Nanoparticles and Hormesis in Complex Adaptive Systems

- Betts JN, Johnson MG, Rygielwicz PT, King GA, and Andersen CP. 2013. Potential for metal contamination by direct sonication of nanoparticle suspensions. *Environ Toxicol Chem* 32:889-893
- Biju V, Itoh T, and Ishikawa M. 2010. Delivering quantum dots to cells: bioconjugated quantum dots for targeted and nonspecific extracellular and intracellular imaging. *Chem Soc Rev* 39:3031-3056
- Bourdineaud JP, Rossignol R, and Brethes D. 2013. Zebrafish: a model animal for analyzing the impact of environmental pollutants on muscle and brain mitochondrial bioenergetics. *Int J Biochem Cell Biol* 45:16-22
- Browning LM, Lee KJ, Huang T, Nallathamby PD, Lowman JE, and Xu XH. 2009. Random walk of single gold nanoparticles in zebrafish embryos leading to stochastic toxic effects on embryonic developments. *Nanoscale* 1:138-152
- Burris JN, Lenaghan SC, Zhang M, and Stewart CN. 2012. Nanoparticle biofabrication using English ivy (*Hedera helix*). *J Nanobiotechnology* 10:41
- Buzea C, Pacheco II, and Robbie K. 2007. Nanomaterials and nanoparticles: sources and toxicity. *Biointerphases* 2:MR17-71
- Calabrese EJ. 2013. Hormetic mechanisms. *Critical Reviews in Toxicology* 43:580-606
- Calabrese EJ, Bachmann KA, Bailer AJ, Bolger PM, Borak J, Cai L, Cedergreen N, Cherian MG, Chiueh CC, Clarkson TW, Cook RR, Diamond DM, Doolittle DJ, Dorato MA, Duke SO, Feinendegen L, Gardner DE, Hart RW, Hastings KL, Hayes AW, Hoffmann GR, Ives JA, Jaworowski Z, Johnson TE, Jonas WB, Kaminski NE, Keller JG, Klaunig JE, Knudsen TB, Kozumbo WJ, Lettieri T, Liu SZ, Maisseu A, Maynard KI, Masoro EJ, McClellan RO, Mehendale HM, Mothersill C, Newlin DB, Nigg HN, Oehme FW, Phalen RF, Philbert MA, Rattan SI, Riviere JE, Rodricks J, Sapolsky RM, Scott BR, Seymour C, Sinclair DA, Smith-Sonneborn J, Snow ET, Spear L, Stevenson DE, Thomas Y, Tubiana M, Williams GM, and Mattson MP. 2007. Biological stress response terminology: Integrating the concepts of adaptive response and preconditioning stress within a hormetic dose-response framework. *Toxicol Appl Pharmacol* 222:122-128
- Calabrese EJ and Mattson MP. 2011. Hormesis provides a generalized quantitative estimate of biological plasticity. *J Cell Commun Signal* 5:25-38
- Cao G and Wang Y. 2011. *Nanostructures and Nanomaterials: Synthesis, Properties, and Applications* 2nd Edition. World Scientific, New Jersey
- Casado-Pascual J, Gomez-Ordóñez J, Morillo M, and Hanggi P. 2003. Subthreshold stochastic resonance: rectangular signals can cause anomalous large gains. *Phys Rev E Stat Nonlin Soft Matter Phys* 68:061104
- Chang X and Vikesland PJ. 2013. Uncontrolled Variability in the Extinction Spectra of C60 Nanoparticle Suspensions. *Langmuir* 29:9685-9693
- Chaowanachan T, Krogstad E, Ball C, and Woodrow KA. 2013. Drug Synergy of Tenofovir and Nanoparticle-Based Antiretrovirals for HIV Prophylaxis. *PLoS One* 8:e61416
- Chen P, Powell BA, Mortimer M, and Ke PC. 2012. Adaptive Interactions between Zinc Oxide Nanoparticles and *Chlorella* sp. *Environ Sci Technol* 46:12178-12185
- Chikramane PS, Kalita D, Suresh AK, Kane SG, and Bellare JR. 2012. Why Extreme Dilutions Reach Non-zero Asymptotes: A Nanoparticulate Hypothesis Based on Froth Flotation. *Langmuir* 28:15864-15875
- Cho GB, Choi SY, Noh JP, Jeon YM, Jung KT, and Nam TH. 2011. Dependence of milling time on electrochemical properties of nano Si electrodes prepared by ball-milling. *J Nanosci Nanotechnol* 11:6262-6265
- Chu SH, Feng DF, Ma YB, and Li ZQ. 2012. Hydroxyapatite nanoparticles inhibit the growth of human glioma cells in vitro and in vivo. *Int J Nanomedicine* 7:3659-3666
- Chudnovsky EM and Friedman JR. 2000. Macroscopic Quantum Coherence in a Magnetic Nanoparticle Above the Surface of a Superconductor. *Physical Review Letters* 85:5206-5209
- Chun YS, Bisht S, Chenna V, Pramanik D, Yoshida T, Hong SM, de Wilde RF, Zhang Z, Huso DL, Zhao M, Rudek MA, Stearns V, Maitra A, and Sukumar S. 2012. Intraductal administration of a polymeric nanoparticle formulation of curcumin (NanoCurc) significantly attenuates incidence of mammary tumors in a rodent chemical carcinogenesis model: Implications for breast cancer chemoprevention in at-risk populations. *Carcinogenesis* 33:2242-2249
- Clark RJ, Dang MK, and Veinot JG. 2010. Exploration of organic acid chain length on water-soluble silicon quantum dot surfaces. *Langmuir* 26:15657-15664
- Clift MJ, Gehr P, and Rothen-Rutishauser B. 2011. Nanotoxicology: a perspective and discussion of whether or not in vitro testing is a valid alternative. *Arch Toxicol* 85:723-731

I. R. Bell and others

- Coffey DS. 1998. Self-organization, complexity, and chaos: the new biology for medicine. *Nature Medicine*. 4:882-885
- Cole AJ, Yang VC, and David AE. 2011. Cancer theranostics: the rise of targeted magnetic nanoparticles. *Trends Biotechnol* 29:323-332
- Costa M, Goldberger AL, and Peng CK. 2002. Multiscale entropy to distinguish physiologic and synthetic RR time series. *Comput Cardiol* 29:137-140
- Costa M, Goldberger AL, and Peng CK. 2005. Multiscale entropy analysis of biological signals. *Physical Review E* 71:1-18
- Costa M, Goldberger AL, and Peng CK. 2002. Multiscale entropy analysis of complex physiologic time series. *Phys Rev Lett* 89:068102:068101-068104
- Costa M, Priplata AA, Lipsitz LA, Wu Z, Huang NE, Goldberger AL, and Peng CK. 2007. Noise and poise: Enhancement of postural complexity in the elderly with a stochastic-resonance-based therapy. *Europhys Lett*. 77:68008
- Cumbo A, Lorber B, Corvini PF, Meier W, and Shahgaldian P. 2013. A synthetic nanomaterial for virus recognition produced by surface imprinting. *Nat Commun* 4:1503
- Czaplicka A, Holyst JA, and Sloot PM. 2013. Noise enhances information transfer in hierarchical networks. *Sci Rep* 3:1223
- Daisy P and Saipriya K. 2012. Biochemical analysis of Cassia fistula aqueous extract and phytochemically synthesized gold nanoparticles as hypoglycemic treatment for diabetes mellitus. *Int J Nanomedicine* 7:1189-1202
- Dar MA, Ingle A, and Rai M. 2013. Enhanced antimicrobial activity of silver nanoparticles synthesized by *Cryphonectria* sp. evaluated singly and in combination with antibiotics. *Nanomedicine* 9:105-110
- Das S, Das J, Samadder A, Bhattacharyya S, Das D, and Khuda-Bukhsh AR. 2013. Biosynthesized silver nanoparticles by ethanolic extracts of *Phytolacca decandra*, *Gelsemium sempervirens*, *Hydrastis canadensis* and *Thuja occidentalis* induce differential cytotoxicity through G2/M arrest in A375 cells. *Colloids and Surfaces B: Biointerfaces* 101:325-336
- Davies PC. 2004. Does quantum mechanics play a non-trivial role in life? *Biosystems* 78:69-79
- DeCastro CL and Mitchell BS. 2002. Nanoparticles from mechanical attrition, in: Baraton MI (Ed.), *Synthesis, Functionalization, and Surface Treatment of Nanoparticles*. American Scientific Publisher, Valencia, CA, pp. 1-15
- Demento SL, Eisenbarth SC, Foellmer HG, Platt C, Caplan MJ, Mark Saltzman W, Mellman I, Ledizet M, Fikrig E, Flavell RA, and Fahmy TM. 2009. Inflammasome-activating nanoparticles as modular systems for optimizing vaccine efficacy. *Vaccine* 27:3013-3021
- Demirovic D and Rattan SI. 2013. Establishing cellular stress response profiles as biomarkers of homeodynamics, health and hormesis. *Exp Gerontol* 48:94-98
- Diwan M, Elamanchili P, Cao M, and Samuel J. 2004. Dose sparing of CpG oligodeoxynucleotide vaccine adjuvants by nanoparticle delivery. *Curr Drug Deliv* 1:405-412
- Du L, Miao X, Jiang Y, Jia H, Tian Q, Shen J, and Liu Y. 2013. An effective strategy for the synthesis of biocompatible gold nanoparticles using danshensu antioxidant: prevention of cytotoxicity via attenuation of free radical formation. *Nanotoxicology* 7:94-300
- European Commission on the Environment. 2011. Nanomaterials, Definition of nanomaterials. Available at http://ec.europa.eu/environment/chemicals/nanotech/faq/definition_en.htm
- Farkas JJ, Korcsmaros T, Kovacs IA, Mihalik A, Palotai R, Simko GI, Szalay KZ, Szalay-Beko M, Vellai T, Wang S, and Csermely P. 2011. Network-based tools for the identification of novel drug targets. *Sci Signal* 4:pt3
- Farmen E, Mikkelsen HN, Evensen O, Einset J, Heier LS, Rosseland BO, Salbu B, Tollefsen KE, and Oughton DH. 2012. Acute and sub-lethal effects in juvenile Atlantic salmon exposed to low mug/L concentrations of Ag nanoparticles. *Aquat Toxicol* 108:78-84
- Foldbjerg R, Irving ES, Hayashi Y, Sutherland D, Thorsen K, Autrup H, and Beer C. 2012. Global gene expression profiling of human lung epithelial cells after exposure to nanosilver. *Toxicol Sci*
- Fredrickson BL and Losada MF. 2005. Positive affect and the complex dynamics of human flourishing. *American Psychologist* 60:678-686
- Garfinkel A, Spano ML, Ditto WL, and Weiss JN. 1992. Controlling cardiac chaos. *Science*. 257:1230-1235
- Gautam S, Dubey P, and Gupta MN. 2013. A facile and green ultrasonic-assisted synthesis of BSA conjugated silver nanoparticles. *Colloids Surf B Biointerfaces* 102:879-883

Nanoparticles and Hormesis in Complex Adaptive Systems

- Geinguenaud F, Souissi I, Fagard R, Motte L, and Lalatonne Y. 2012. Electrostatic assembly of a DNA superparamagnetic nano-tool for simultaneous intracellular delivery and in situ monitoring. *Nanomedicine* 8:1106-1115
- Ghosh D, Choudhury ST, Ghosh S, Mandal AK, Sarkar S, Ghosh A, Saha KD, and Das N. 2012. Nanocapsulated curcumin: oral chemopreventive formulation against diethylnitrosamine induced hepatocellular carcinoma in rat. *Chem Biol Interact* 195:206-214
- Goldberger AL. 1996. Non-linear dynamics for clinicians: chaos theory, fractals, and complexity at the bedside. *Lancet* 347:1312-1314
- Goldberger AL, Peng CK, and Lipsitz LA. 2002. What is physiologic complexity and how does it change with aging and disease? *Neurobiol Aging* 23:23-26
- Gosnell BA. 2005. Sucrose intake enhances behavioral sensitization produced by cocaine. *Brain Res* 1031:194-201
- Gualtieri M, Skuland T, Iversen TG, Lag M, Schwarze P, Bilanicova D, Pojana G, and Refsnes M. 2012. Importance of agglomeration state and exposure conditions for uptake and pro-inflammatory responses to amorphous silica nanoparticles in bronchial epithelial cells. *Nanotoxicology* 6:700-712
- Gupta A and Wiggers H. 2011. Freestanding silicon quantum dots: origin of red and blue luminescence. *Nanotechnology* 22:055707
- Hale HB. 1969. Cross-adaptation. *Environmental Research* 2:423-434
- Hannah DC, Yang J, Podsiadlo P, Chan MK, Demortiere A, Gosztola DJ, Prakapenka VB, Schatz GC, Kortshagen U, and Schaller RD. 2012. On the origin of photoluminescence in silicon nanocrystals: pressure-dependent structural and optical studies. *Nano Lett* 12:4200-4205
- Harhaji L, Isakovic A, Raicevic N, Markovic Z, Todorovic-Markovic B, Nikolic N, Vranjes-Djuric S, Markovic I, and Trajkovic V. 2007. Multiple mechanisms underlying the anticancer action of nanocrystalline fullerene. *Eur J Pharmacol* 568:89-98
- Hassan TA, Rangari VK, Rana RK, and Jeelani S. 2013. Sonochemical effect on size reduction of CaCO₃ nanoparticles derived from waste eggshells. *Ultrason Sonochem* 20:1308-1315
- Hatef A, Sadeghi SM, and Singh MR. 2012. Coherent molecular resonances in quantum dot-metallic nanoparticle systems: coherent self-renormalization and structural effects. *Nanotechnology* 23:205203
- Ho D, Sun X, and Sun S. 2011. Monodisperse magnetic nanoparticles for theranostic applications. *Acc Chem Res* 44:875-882
- Ho YP and Leong KW. 2010. Quantum dot-based theranostics. *Nanoscale* 2:60-68
- Hollenstein T. 2007. State space grids: analyzing dynamics across development. *International Journal of Behavioral Development*. 31:384-396
- Hossu M, Ma L, and Chen W. 2010. Nonlinear enhancement of spontaneous biophoton emission of sweet potato by silver nanoparticles. *J Photochem Photobiol B*. 99:44-48
- Huang T, Browning LM, and Xu XH. 2012. Far-field photostable optical nanoscopy (PHOTON) for real-time super-resolution single-molecular imaging of signaling pathways of single live cells. *Nanoscale* 4:2797-2812
- Huang X, Kang B, Qian W, Mackey MA, Chen PC, Oyelere AK, El-Sayed IH, and El-Sayed MA. 2010. Comparative study of photothermolysis of cancer cells with nuclear-targeted or cytoplasm-targeted gold nanospheres: continuous wave or pulsed lasers. *J Biomed Opt* 15:058002
- Hudecova A, Kusznierevicz B, Runden-Pran E, Magdolenova Z, Hasplova K, Rinna A, Fjellsbo LM, Kruszewski M, Lankoff A, Sandberg WJ, Refsnes M, Skuland T, Schwarze P, Brunborg G, Bjaras M, Collins A, Miadokova E, Galova E, and Dusinska M. 2012. Silver nanoparticles induce pre-mutagenic DNA oxidation that can be prevented by phytochemicals from *Gentiana asclepiadea*. *Mutagenesis* 27:759-769
- Iavicoli I, Calabrese EJ, and Nascarella MA. 2010. Exposure to nanoparticles and hormesis. *Dose Response* 8:501-517
- Ichiki A and Tadokoro Y. 2013. Relation between optimal nonlinearity and non-Gaussian noise: enhancing a weak signal in a nonlinear system. *Phys Rev E Stat Nonlin Soft Matter Phys* 87:012124
- Im YB, Wahab R, Ameen S, Kim YS, Yang OB, and Shin HS. 2011. Synthesis and characterization of high-purity silica nanosphere from rice husk. *J Nanosci Nanotechnol* 11:5934-5938
- International Organization for Standardization. 2005. ISO/TC 229 Nanotechnologies, Standardization in the field of nanotechnologies. Available at http://www.iso.org/iso/iso_technical_committee?commid=381983

I. R. Bell and others

- Isoda K, Hasezaki T, Kondoh M, Tsutsumi Y, and Yagi K. 2011. Effect of surface charge on nano-sized silica particles-induced liver injury. *Pharmazie* 66:278-281
- Ives JA, Moffett JR, Arun P, Lam D, Todorov TI, Brothers AB, Anick DJ, Centeno J, Namboodiri MA, and Jonas WB. 2010. Enzyme stabilization by glass-derived silicates in glass-exposed aqueous solutions. *Homeopathy* 99:15-24
- Jena P, Mohanty S, Mallick R, Jacob B, and Sonawane A. 2012. Toxicity and antibacterial assessment of chitosancoated silver nanoparticles on human pathogens and macrophage cells. *Int J Nanomedicine* 7:1805-1818
- Jiang W, Kim BY, Rutka JT, and Chan WC. 2008. Nanoparticle-mediated cellular response is size-dependent. *Nat Nanotechnol* 3:145-150
- Jonasson S, Gustafsson A, Koch B, and Bucht A. 2013. Inhalation exposure of nano-scaled titanium dioxide (TiO₂) particles alters the inflammatory responses in asthmatic mice. *Inhal Toxicol* 25:179-191
- Joshi MD and Muller RH. 2009. Lipid nanoparticles for parenteral delivery of actives. *Eur J Pharm Biopharm* 71:161-172
- Ju-Nam Y and Lead JR. 2008. Manufactured nanoparticles: an overview of their chemistry, interactions and potential environmental implications. *Sci Total Environ* 400:396-414
- Kang Z, Liu Y, and Lee ST. 2011. Small-sized silicon nanoparticles: new nanolights and nanocatalysts. *Nanoscale* 3:777-791
- Kaur J and Tikoo K. 2012. Evaluating cell specific cytotoxicity of differentially charged silver nanoparticles. *Food Chem Toxicol* 51C:1-14
- Kelty-Stephen DG and Dixon JA. 2013. Temporal correlations in postural sway moderate effects of stochastic resonance on postural stability. *Hum Mov Sci* 32:91-105
- Kiel S, Grinberg O, Perkas N, Charmet J, Kepner H, and Gedanken A. 2012. Forming nanoparticles of water-soluble ionic molecules and embedding them into polymer and glass substrates. *Beilstein J Nanotechnol* 3:267-276
- Kim TH, Kim M, Park HS, Shin US, Gong MS, and Kim HW. 2012. Size-dependent cellular toxicity of silver nanoparticles. *J Biomed Mater Res A* 100:1033-1043
- Kleps I, Ignat T, Miu M, Craciunoiu F, Trif M, Simion M, Bragaru A, and Dinescu A. 2010. Nanostructured silicon particles for medical applications. *J Nanosci Nanotechnol* 10:2694-2700
- Koning GA and Krijger GC. 2007. Targeted multifunctional lipid-based nanocarriers for image-guided drug delivery. *Anticancer Agents Med Chem* 7:425-440
- Korn H and Faure P. 2003. Is there chaos in the brain? II. Experimental evidence and related models. [Review] [343 refs]. *Comptes Rendus Biologies* 326:787-840
- Krawiecki A, Sukiennicki A, and Kosinski RA. 2000. Stochastic resonance and noise-enhanced order with spatiotemporal periodic signal. *Phys Rev E Stat Phys Plasmas Fluids Relat Interdiscip Topics* 62:7683-7689
- Kumar V, Kumari A, Guleria P, and Yadav SK. 2012. Evaluating the toxicity of selected types of nanochemicals. *Rev Environ Contam Toxicol* 215:39-121
- Lagisz M, Hector KL, and Nakagawa S. 2013. Life extension after heat shock exposure: Assessing meta-analytic evidence for hormesis. *Ageing Res Rev* 12:653-60
- Lanao JM, Briones E, and Colino CI. 2007. Recent advances in delivery systems for anti-HIV1 therapy. *J Drug Target* 15:21-36
- Lankoff A, Arabski M, Wegierek-Ciuk A, Kruszewski M, Lisowska H, Banasik-Nowak A, Rozga-Wijas K, Wojewodzka M, and Slomkowski S. 2013. Effect of surface modification of silica nanoparticles on toxicity and cellular uptake by human peripheral blood lymphocytes in vitro. *Nanotoxicology* 7:235-50
- Launay JC, Besnard Y, Guinet-Lebreton A, and Savourey G. 2006. Acclimation to intermittent hypobaric hypoxia modifies responses to cold at sea level. *Aviat Space Environ Med* 77:1230-1235
- Lee CW, Yen FL, Huang HW, Wu TH, Ko HH, Tzeng WS, and Lin CC. 2012a. Resveratrol Nanoparticle System Improves Dissolution Properties and Enhances the Hepatoprotective Effect of Resveratrol through Antioxidant and Anti-Inflammatory Pathways. *J Agric Food Chem* 60:4662-4671
- Lee CY, Choi W, Han JH, and Strano MS. 2010. Coherence resonance in a single-walled carbon nanotube ion channel. *Science* 329:1320-1324
- Lee I, Liu X, Chongwu Z, and Kosko B. 2006. Noise-Enhanced Detection of Subthreshold Signals With Carbon Nanotubes. *Nanotechnology, IEEE Transactions on* 5:613-627

Nanoparticles and Hormesis in Complex Adaptive Systems

- Lee KJ, Browning LM, Nallathamby PD, Desai T, Cherukuri PK, and Xu XH. 2012b. In vivo quantitative study of sized-dependent transport and toxicity of single silver nanoparticles using zebrafish embryos. *Chem Res Toxicol* 25:1029-1046
- Lenaghan SC, Burris JN, Chourey K, Huang Y, Xia L, Lady B, Sharma R, Pan C, Lejeune Z, Foister S, Hettich RL, Stewart CN, Jr., and Zhang M. 2013. Isolation and chemical analysis of nanoparticles from English ivy (*Hedera helix* L.). *J R Soc Interface* 10:20130392
- Lim DH, Jang J, Kim S, Kang T, Lee K, and Choi IH. 2012. The effects of sub-lethal concentrations of silver nanoparticles on inflammatory and stress genes in human macrophages using cDNA microarray analysis. *Biomaterials* 33:4690-4699
- Lindsay SM. 2010. Introduction to Nanoscience. Oxford
- Liu D, Wu Q, Chen H, and Chang PR. 2009. Transitional properties of starch colloid with particle size reduction from micro- to nanometer. *J Colloid Interface Sci* 339:117-124
- Liu L, Randolph TW, and Carpenter JF. 2012. Particles shed from syringe filters and their effects on agitation-induced protein aggregation. *J Pharm Sci* 101:2952-2959
- Liu Y, Kathan K, Saad W, and Prudhomme RK. 2007. Ostwald ripening of B-carotene nanoparticles. *Physical Review Letters* 98:1-4
- Lloyd S. 2011. Quantum coherence in biological systems. *Journal of Physics: Conference Series International Symposium "Nanoscience and Quantum Physics 2011"*:012037
- Losada M. 1999. The complex dynamics of high performance teams. *Mathematical Computer Modelling* 30:179-192
- Losada M and Heaphy E. 2004. The role of positivity and connectivity in the performance of business teams: a nonlinear dynamics model. *American Behavioral Scientist* 47:740-765
- Lu X, Tian Y, Zhao Q, Jin T, Xiao S, and Fan X. 2011. Integrated metabonomics analysis of the size-response relationship of silica nanoparticles-induced toxicity in mice. *Nanotechnology* 22:055101
- Lunt HC, Barwood MJ, Corbett J, and Tipton MJ. 2010. 'Cross-adaptation': habituation to short repeated cold-water immersions affects the response to acute hypoxia in humans. *J Physiol* 588:3605-3613
- Luther JM, Jain PK, Ewers T, and Alivisatos AP. 2011. Localized surface plasmon resonances arising from free carriers in doped quantum dots. *Nat Mater* 10:361-366
- Magalhaes FH and Kohn AF. 2011. Vibratory noise to the fingertip enhances balance improvement associated with light touch. *Experimental brain research. Experimentelle Hirnforschung. Experimentation cerebrale* 209:139-151
- Mahony D, Cavallaro AS, Stahr F, Mahony TJ, Qiao SZ, and Mitter N. 2013. Mesoporous Silica Nanoparticles Act as a Self-Adjuvant for Ovalbumin Model Antigen in Mice. *Small* 9:3138-46
- Malarczyk E, Pazdzioch-Czochra M, Graz M, Kochmanska-Rdest J, and Jarosz-Wilkolazka A. 2011. Nonlinear changes in the activity of the oxygen-dependent demethylase system in *Rhodococcus erythropolis* cells in the presence of low and very low doses of formaldehyde. *Nonlinear Biomed Phys* 5:9
- McDonnell MD and Abbott D. 2009. What Is Stochastic Resonance? Definitions, Misconceptions, Debates, and Its Relevance to Biology. *PLoS Comput Biol* 5:e1000348
- McGuinness LP, Yan Y, Stacey A, Simpson DA, Hall LT, Maclaurin D, Prawer S, Mulvaney P, Wrachtrup J, Caruso F, Scholten RE, and Hollenberg LC. 2011. Quantum measurement and orientation tracking of fluorescent nanodiamonds inside living cells. *Nat Nanotechnol* 6:358-363
- McKibbin SR, Scappucci G, Pok W, and Simmons MY. 2013. Epitaxial top-gated atomic-scale silicon wire in a three-dimensional architecture. *Nanotechnology* 24:045303
- Melancon MP, Zhou M, and Li C. 2011. Cancer theranostics with near-infrared light-activatable multimodal nanoparticles. *Acc Chem Res* 44:947-956
- Merisko-Liversidge E and Liversidge GG. 2011. Nanosizing for oral and parenteral drug delivery: a perspective on formulating poorly-water soluble compounds using wet media milling technology. *Adv Drug Deliv Rev* 63:427-440
- Mihalik A and Csermely P. 2011. Heat shock partially dissociates the overlapping modules of the yeast protein-protein interaction network: a systems level model of adaptation. *PLoS Comput Biol* 7:e1002187
- Milisav I, Poljsak B, and Suput D. 2012. Adaptive response, evidence of cross-resistance and its potential clinical use. *Int J Mol Sci* 13:10771-10806
- Mittal AK, Chisti Y, and Banerjee UC. 2013. Synthesis of metallic nanoparticles using plant extracts. *Biotechnol Adv* 31:346-56

I. R. Bell and others

- Moss F, Ward LM, and Sannita WG. 2004. Stochastic resonance and sensory information processing: a tutorial and review of application. *Clin Neurophysiol* 115:267-281
- Moulari B, Beduneau A, Pellequer Y, and Lamprecht A. 2013. Nanoparticle targeting to inflamed tissues of the gastrointestinal tract. *Curr Drug Deliv* 10:9-17
- Mudunkotuwa IA and Grassian VH. 2011. The devil is in the details (or the surface): impact of surface structure and surface energetics on understanding the behavior of nanomaterials in the environment. *J Environ Monit* 13:1135-1144
- Murdock RC, Braydich-Stolle L, Schrand AM, Schlager JJ, and Hussain SM. 2008. Characterization of nanomaterial dispersion in solution prior to in vitro exposure using dynamic light scattering technique. *Toxicol Sci* 101:239-253
- Nascarella MA and Calabrese EJ. 2012. A method to evaluate hormesis in nanoparticle dose-responses. *Dose Response* 10:344-354
- Ning XH and Chen SH. 2006. Mild hypothermic cross adaptation resists hypoxic injury in hearts: a brief review. *Chin J Physiol* 49:213-222
- Osborne OJ, Johnston BD, Moger J, Balousha M, Lead JR, Kudoh T, and Tyler CR. 2012. Effects of particle size and coating on nanoscale Ag and TiO₂ exposure in zebrafish (*Danio rerio*) embryos. *Nanotoxicology* 7:1315-1324
- Oyewumi MO, Kumar A, and Cui Z. 2010. Nano-microparticles as immune adjuvants: correlating particle sizes and the resultant immune responses. *Expert Rev Vaccines* 9:1095-1107
- Pandey S, Thakur M, Shah R, Oza G, Mewada A, and Sharon M. 2013. A comparative study of economical separation and aggregation properties of biologically capped and thiol functionalized gold nanoparticles: Selecting the eco-friendly trojan horses for biological applications. *Colloids Surf B Biointerfaces* 109:25-31
- Park Y, Noh HJ, Han L, Kim HS, Kim YJ, Choi JS, Kim CK, Kim YS, and Cho S. 2012. *Artemisia capillaris* extracts as a green factory for the synthesis of silver nanoparticles with antibacterial activities. *J Nanosci Nanotechnol* 12:7087-7095
- Petkar KC, Chavhan SS, Agatonovik-Kustrin S, and Sawant KK. 2011. Nanostructured materials in drug and gene delivery: a review of the state of the art. *Crit Rev Ther Drug Carrier Syst* 28:101-164
- Pham KN, Fullston D, and Sagoe-Crentsil K. 2007. Surface modification for stability of nano-sized silica colloids. *J Colloid Interface Sci* 315:123-127
- Pickering AM, Vojtovich L, Tower J, and KJ AD. 2013. Oxidative stress adaptation with acute, chronic, and repeated stress. *Free Radic Biol Med* 55:109-118
- Pinamonti G, Marro J, and Torres JJ. 2012. Stochastic resonance crossovers in complex networks. *PLoS One* 7:e51170
- Pincus D and Metten A. 2010. Nonlinear dynamics in biopsychosocial resilience. *Nonlinear Dynamics Psychol Life Sci* 14:353-380
- Prakash DJ, Arulkumar S, and Sabesan M. 2010. Effect of nanohypericum (*Hypericum perforatum* gold nanoparticles) treatment on restraint stress induced behavioral and biochemical alteration in male albino mice. *Pharmacognosy Res* 2:330-334
- Raja WK, Satti J, Liu G, and Castracane J. 2013. Dose Response of MTLn3 Cells to Serial Dilutions of Arsenic Trioxide and Ionizing Radiation. *Dose Response* 11:29-40
- Rao KS, El-Hami K, Kodaki T, Matsushige K, and Makino K. 2005. A novel method for synthesis of silica nanoparticles. *J Colloid Interface Sci* 289:125-131
- Richert LE, Servid AE, Harmsen AL, Rynda-Apple A, Han S, Wiley JA, Douglas T, and Harmsen AG. 2012. A virus-like particle vaccine platform elicits heightened and hastened local lung mucosal antibody production after a single dose. *Vaccine* 30:3653-3665
- Ristorcelli E, Beraud E, Mathieu S, Lombardo D, and Verine A. 2009. Essential role of Notch signaling in apoptosis of human pancreatic tumoral cells mediated by exosomal nanoparticles. *Int J Cancer* 125:1016-1026
- Rodrigues MT, Ajayan PM, and Silva GG. 2013. Fast vortex-assisted self-assembly of carbon nanoparticles on an air-water interface. *J Phys Chem B* 117:6524-6533
- Rodriguez M, Snoek LB, Riksen JA, Bevers RP, and Kammenga JE. 2012. Genetic variation for stress-response hormesis in *C. elegans* lifespan. *Exp Gerontol* 47:581-587
- Roduner E. 2006. Size matters: why nanomaterials are different. *Chemical Society Reviews* 35:583-592
- Rowe DJ, Jeong JS, Mkhoyan KA, and Kortshagen UR. 2013. Phosphorus-doped silicon nanocrystals exhibiting mid-infrared localized surface plasmon resonance. *Nano Lett* 13:1317-1322

Nanoparticles and Hormesis in Complex Adaptive Systems

- Ruan B and Jacobi M. 2012. Ultrasonication effects on thermal and rheological properties of carbon nanotube suspensions. *Nanoscale Research Letters* 7:127
- Salavati-Niasari M, Javidi J, and Dadkhah M. 2012. Ball milling synthesis of silica nanoparticle from rice husk ash for drug delivery application. *Comb Chem High Throughput Screen* 16:458-62
- Sandberg WJ, Lag M, Holme JA, Friede B, Gualtieri M, Kruszewski M, Schwarze PE, Skuland T, and Refsnes M. 2012. Comparison of non-crystalline silica nanoparticles in IL-1 β release from macrophages. *Part Fibre Toxicol* 9:32
- Sayed D, Al-Sadoon MK, and Badr G. 2012. Silica Nanoparticles Sensitize Human Multiple Myeloma Cells to Snake (*Walterinnesia aegyptia*) Venom-Induced Apoptosis and Growth Arrest. *Oxidative medicine and cellular longevity* 2012:386286
- Schiff SJ, Jerger K, Duong DH, Chang T, Spano ML, and Ditto WL. 1994. Controlling chaos in the brain. *Nature* 370:615-620
- Shalchian M, Grisolia J, Assayag GB, Coffin H, Atarodi SM, and Claverie A. 2005. Room-temperature quantum effect in silicon nanoparticles obtained by low-energy ion implantation and embedded in a nanometer scale capacitor. *Applied Physics Letters* 86:163111-163113
- Shannahan JH, Kodavanti UP, and Brown JM. 2012. Manufactured and airborne nanoparticle cardiopulmonary interactions: a review of mechanisms and the possible contribution of mast cells. *Inhal Toxicol* 24:320-339
- Shi Y, Zhang JH, Jiang M, Zhu LH, Tan HQ, and Lu B. 2010a. Synergistic genotoxicity caused by low concentration of titanium dioxide nanoparticles and p,p'-DDT in human hepatocytes. *Environ Mol Mutagen* 51:192-204
- Shi Z, Huang X, Liu B, Tao H, Cai Y, and Tang R. 2010b. Biological response of osteosarcoma cells to size-controlled nanostructured hydroxyapatite. *J Biomater Appl* 25:19-37
- Shirali AC, Look M, Du W, Kassis E, Stout-Delgado HW, Fahmy TM, and Goldstein DR. 2011. Nanoparticle delivery of mycophenolic acid upregulates PD-L1 on dendritic cells to prolong murine allograft survival. *American journal of transplantation : official journal of the American Society of Transplantation and the American Society of Transplant Surgeons* 11:2582-2592
- Siddiqi NJ, Abdelhalim MA, El-Ansary AK, Alhomida AS, and Ong WY. 2012. Identification of potential biomarkers of gold nanoparticle toxicity in rat brains. *J Neuroinflammation* 9:123
- Singh DK, Jagannathan R, Khandelwal P, Abraham PM, and Poddar P. 2013. In situ synthesis and surface functionalization of gold nanoparticles with curcumin and their antioxidant properties: an experimental and density functional theory investigation. *Nanoscale* 5:1882-1893
- Snitka V, Naumenko DO, Ramanauskaite L, Kravchenko SA, and Snopok BA. 2012. Generation of diversiform gold nanostructures inspired by honey's components: Growth mechanism, characterization, and shape separation by the centrifugation-assisted sedimentation. *J Colloid Interface Sci* 386:99-106
- Song L, Yang K, Jiang W, Du P, and Xing B. 2012. Adsorption of bovine serum albumin on nano and bulk oxide particles in deionized water. *Colloids Surf B Biointerfaces* 94:341-346
- Sorg BA, Tschirgi ML, Swindell S, Chen L, and Fang J. 2001. Repeated formaldehyde effects in an animal model for multiple chemical sensitivity. *Annals of the New York Academy of Sciences*. 933:57-67
- Sorg BA, Willis JR, See RE, Hopkins B, and Westberg HH. 1998. Repeated low-level formaldehyde exposure produces cross-sensitization to cocaine: possible relevance to chemical sensitivity in humans. *Neuropsychopharmacology* 18:385-394
- Soti C and Csermely P. 2003. Aging and molecular chaperones. *Exp Gerontol* 38:1037-1040
- Soti C and Csermely P. 2006. Pharmacological modulation of the heat shock response. *Handb Exp Pharmacol*:417-436
- Soti C and Csermely P. 2007. Aging cellular networks: chaperones as major participants. *Exp Gerontol* 42:113-119
- Soumya RS, Ghosh S, and Abraham ET. 2010. Preparation and characterization of guar gum nanoparticles. *Int J Biol Macromol* 46:267-269
- Stark M. 2012. The sandpile model: optimal stress and hormesis. *Dose Response* 10:66-74
- Stark WJ. 2011. Nanoparticles in biological systems. *Angew Chem Int Ed Engl* 50:1242-1258
- Stovbun SV, Kiselev AV, Zanin AM, Kalinina TS, Voronina TA, Mikhailov AI, and Berlin AA. 2012. Effects of physicochemical forms of phenazepam and Panavir on their action at ultra-low doses. *Bull Exp Biol Med* 153:455-458

I. R. Bell and others

- Sugarman J, Tsai S, Santamaria P, and Khadra A. 2013. Quantifying the importance of pMHC valency, total pMHC dose and frequency on nanoparticle therapeutic efficacy. *Immunol Cell Biol* 91:350-359
- Sun H, Chen X, Chen D, Dong M, Fu X, Li Q, Liu X, Wu Q, Qiu T, Wan T, and Li S. 2012. Influences of surface coatings and components of FePt nanoparticles on the suppression of glioma cell proliferation. *Int J Nanomedicine* 7:3295-3307
- Sur I, Altunbek M, Kahraman M, and Culha M. 2012. The influence of the surface chemistry of silver nanoparticles on cell death. *Nanotechnology* 23:375102
- Sur I, Cam D, Kahraman M, Baysal A, and Culha M. 2010. Interaction of multi-functional silver nanoparticles with living cells. *Nanotechnology* 21:175104
- Suriyakalaa U, Antony JJ, Suganya S, Siva D, Sukirtha R, Kamalakkannan S, Pichiah PB, and Achiraman S. 2012. Hepatocurative activity of biosynthesized silver nanoparticles fabricated using *Andrographis paniculata*. *Colloids Surf B Biointerfaces* 102C:189-194
- Szalay MS, Kovacs IA, Korcsmaros T, Bode C, and Csermely P. 2007. Stress-induced rearrangements of cellular networks: Consequences for protection and drug design. *FEBS Lett* 581:3675-3680
- Tang C, Zhou T, Yang J, Zhang Q, Chen F, Fu Q, and Yang L. 2011. Wet-grinding assisted ultrasonic dispersion of pristine multi-walled carbon nanotubes (MWCNTs) in chitosan solution. *Colloids Surf B Biointerfaces* 86:189-197
- Tang M, Zhang T, Xue Y, Wang S, Huang M, Yang Y, Lu M, Lei H, Kong L, and Yuepu P. 2010. Dose dependent in vivo metabolic characteristics of titanium dioxide nanoparticles. *J Nanosci Nanotechnol* 10:8575-8583
- Tantra R, Tompkins J, and Quincey P. 2010. Characterisation of the de-agglomeration effects of bovine serum albumin on nanoparticles in aqueous suspension. *Colloids Surf B Biointerfaces* 75:275-281
- Thurber A, Wingett DG, Rasmussen JW, Layne J, Johnson L, Tenne DA, Zhang J, Hanna CB, and Punnoose A. 2012. Improving the selective cancer killing ability of ZnO nanoparticles using Fe doping. *Nanotoxicology* 6:440-452
- Torres JL and Ruiz MAG. 1996. Stochastic resonance and the homeopathic effect. *British Homoeopathic Journal* 85(3):134-140
- Tournebise J, Boudier A, Joubert O, Eidi H, Bartosz G, Maincent P, Leroy P, and Sapin-Minet A. 2012. Impact of gold nanoparticle coating on redox homeostasis. *Int J Pharm* 438:107-116
- Tripathi A, Chandrasekaran N, Raichur AM, and Mukherjee A. 2009. Antibacterial applications of silver nanoparticles synthesized by aqueous extract of *Azadirachta indica* (Neem) leaves. *J Biomed Nanotechnol* 5:93-98
- Troia A, Giovannozzi A, and Amato G. 2009. Preparation of tunable silicon q-dots through ultrasound. *Ultrason Sonochem* 16:448-451
- Truong L, Tilton SC, Zaikova T, Richman E, Waters KM, Hutchison JE, and Tanguay RL. 2013. Surface functionalities of gold nanoparticles impact embryonic gene expression responses. *Nanotoxicology* 7:192-201
- Umashankari J, Inbakandan D, Ajithkumar TT, and Balasubramanian T. 2012. Mangrove plant, *Rhizophora mucronata* (Lamk, 1804) mediated one pot green synthesis of silver nanoparticles and its antibacterial activity against aquatic pathogens. *Aquatic biosystems* 8:11
- Vaiserman AM. 2010. Hormesis, adaptive epigenetic reorganization, and implications for human health and longevity. *Dose Response*. 8:16-21
- Vaiserman AM. 2011. Hormesis and epigenetics: is there a link? *Ageing Res Rev* 10:413-421
- Van Hoecke K, De Schamphelaere KA, Ramirez-Garcia S, Van der Meer P, Smagghe G, and Janssen CR. 2011. Influence of alumina coating on characteristics and effects of SiO₂ nanoparticles in algal growth inhibition assays at various pH and organic matter contents. *Environ Int* 37:1118-1125
- Van Hoecke K, De Schamphelaere KA, Van der Meer P, Lucas S, and Janssen CR. 2008. Ecotoxicity of silica nanoparticles to the green alga *Pseudokirchneriella subcapitata*: importance of surface area. *Environ Toxicol Chem* 27:1948-1957
- Van Wijk R and Wiegant FA. 2010. Postconditioning hormesis and the homeopathic Similia principle: molecular aspects. *Hum Exp Toxicol* 29:561-565
- Van Wijk R and Wiegant FA. 2011. Postconditioning hormesis and the similia principle. *Front Biosci (Elite Ed)* 3:1128-1138
- Vesterdal LK, Folkmann JK, Jacobsen NR, Sheykhzade M, Wallin H, Loft S, and Moller P. 2010. Pulmonary exposure to carbon black nanoparticles and vascular effects. *Part Fibre Toxicol* 7:33

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- Vijayaraghavan K and Nalini SP. 2010. Biotemplates in the green synthesis of silver nanoparticles. *Biotechnol J* 5:1098-1110
- Vivero-Escoto JL, Slowing, II, Trewyn BG, and Lin VS. 2010. Mesoporous silica nanoparticles for intracellular controlled drug delivery. *Small* 6:1952-1967
- Wang M and Thanou M. 2010. Targeting nanoparticles to cancer. *Pharmacol Res* 62:90-99
- Wang Q, Bao Y, Ahire J, and Chao Y. 2012a. Co-encapsulation of Biodegradable Nanoparticles with Silicon Quantum Dots and Quercetin for Monitored Delivery. *Advanced healthcare materials* 2:459-66
- Wang T, Jiang H, Zhao Q, Wang S, Zou M, and Cheng G. 2012b. Enhanced mucosal and systemic immune responses obtained by porous silica nanoparticles used as an oral vaccine adjuvant: Effect of silica architecture on immunological properties. *Int J Pharm* 436:351-358
- Wang W, Luo M, Fu Y, Wang S, Efferth T, and Zu Y. 2013. Glycyrrhizic acid nanoparticles inhibit LPS-induced inflammatory mediators in 264.7 mouse macrophages compared with unprocessed glycyrrhizic acid. *Int J Nanomedicine* 8:1377-1383
- Wang Y and Chen L. 2011. Quantum dots, lighting up the research and development of nanomedicine. *Nanomedicine* 7:385-402
- Wiegant FA, Prins HA, and Van Wijk R. 2011. Postconditioning hormesis put in perspective: an overview of experimental and clinical studies. *Dose Response* 9:209-224
- Wiegant FA, Spiekier N, and van Wijk R. 1998. Stressor-specific enhancement of hsp induction by low doses of stressors in conditions of self- and cross-sensitization. *Toxicology* 127:107-119
- Winnik FM and Maysinger D. 2013. Quantum Dot Cytotoxicity and Ways To Reduce It. *Acc Chem Res* 46:672-680
- Wong TW. 2011. Oral fast-release solid dispersion-paradigm shift to nanoparticles. *Recent Pat Drug Deliv Formul* 5:227-243
- Xia Y, Xiong Y, Lim B, and Skrabalak SE. 2009. Shape-controlled synthesis of metal nanocrystals: simple chemistry meets complex physics? *Angew Chem Int Ed Engl* 48:60-103
- Yang Y, Yang AL, Yang RQ, Yuan GJ, and Shi YL. 2011. Investigation of the enhancement fluorescence of ethanol doped SiO₂ nanoparticles. *J Nanosci Nanotechnol* 11:9717-9720
- Yao P and Hughes S. 2009. Macroscopic entanglement and violation of Bell's inequalities between two spatially separated quantum dots in a planar photonic crystal system. *Opt Express* 17:11505-11514
- Yen FL, Wu TH, Lin LT, Cham TM, and Lin CC. 2008. Nanoparticles formulation of *Cuscuta chinensis* prevents acetaminophen-induced hepatotoxicity in rats. *Food Chem Toxicol* 46:1771-1777
- Yoo D, Lee JH, Shin TH, and Cheon J. 2011. Theranostic magnetic nanoparticles. *Acc Chem Res* 44:863-874
- Yoo JW, Yun DS, and Kim HJ. 2006. Influence of reaction parameters on size and shape of silica nanoparticles. *J Nanosci Nanotechnol* 6:3343-3346
- Yu H, Wang J, Du J, Deng B, Wei X, and Liu C. 2013. Effects of time delay on the stochastic resonance in small-world neuronal networks. *Chaos* 23:013128
- Zhang H, He X, Zhang Z, Zhang P, Li Y, Ma Y, Kuang Y, Zhao Y, and Chai Z. 2011. Nano-CeO₂ exhibits adverse effects at environmental relevant concentrations. *Environ Sci Technol* 45:3725-3730
- Zhang S, Chen CS, Schwehr K, Quigg A, Chin WC, Santschi PH, Spurgin J, and Jiang Y. 2012. Aggregation, Dissolution and Stability of Quantum Dots in Marine Environments: The Importance of Extracellular Polymeric Substances. *Environ Sci Technol* 46:8764-72
- Zhao L and Feng SS. 2010. Enhanced oral bioavailability of paclitaxel formulated in vitamin E-TPGS emulsified nanoparticles of biodegradable polymers: in vitro and in vivo studies. *J Pharm Sci* 99:3552-3560
- Zhao L, Peng B, Hernandez-Viezcas JA, Rico C, Sun Y, Peralta-Videa JR, Tang X, Niu G, Jin L, Varela-Ramirez A, Zhang JY, and Gardea-Torresdey JL. 2012. Stress Response and Tolerance of *Zea mays* to CeO(2) Nanoparticles: Cross Talk among H(2)O(2), Heat Shock Protein, and Lipid Peroxidation. *ACS Nano* 6:9615-22
- Zhou H and Lee J. 2011. Nanoscale hydroxyapatite particles for bone tissue engineering. *Acta Biomater* 7:2769-2781
- Zhu M, Li Y, Shi J, Feng W, Nie G, and Zhao Y. 2012a. Exosomes as extrapulmonary signaling conveyors for nanoparticle-induced systemic immune activation. *Small* 8:404-412

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- Zhu M, Tian X, Song X, Li Y, Tian Y, Zhao Y, and Nie G. 2012b. Nanoparticle-Induced Exosomes Target Antigen-Presenting Cells to Initiate Th1-Type Immune Activation. *Small* 8:2841-2848
- Zhu S, Oberdorster E, and Haasch ML. 2006. Toxicity of an engineered nanoparticle (fullerene, C60) in two aquatic species, *Daphnia* and fathead minnow. *Mar Environ Res* 62 Suppl:S5-9