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INCREASED RADIORESISTANCE TO LETHAL DOSES OF GAMMA RAYS IN MICE AND RATS AFTER EXPOSURE TO MICROWAVE RADIATION EMITTED BY A GSM MOBILE PHONE SIMULATOR

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□ The aim of this study was to investigate the effect of pre-irradiation with microwaves on the induction of radioadaptive response. In the 1st phase of the study, 110 male mice were divided into 8 groups. The animals in these groups were exposed/sham-exposed to microwave, low dose rate gamma or both for 5 days. On day six, the animals were exposed to a lethal dose (LD). In the 2nd phase, 30 male rats were divided into 2 groups of 15 animals. The 1st group received microwave exposure. The 2nd group (controls) received the same LD but there was no treatment before the LD. On day 5, all animals were whole-body irradiated with the LD. Statistically significant differences between the survival rate of the mice only exposed to lethal dose of gamma radiation before irradiation with a lethal dose of gamma radiation with those of the animals pre-exposed to either microwave ($p=0.02$), low dose rate gamma ($p=0.001$) or both of these physical adapting doses ($p=0.003$) were observed. Likewise, a statistically significant difference between survival rates of the rats in control and test groups was observed. Altogether, these experiments showed that exposure to microwave radiation may induce a significant survival adaptive response.

Keywords: Microwave, Adaptive Response, Nonionizing Radiation, Survival, GSM Mobile Phone

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INTRODUCTION

Radioadaptive response or radiation-induced adaptive response is defined as the acquisition of radiation resistance against irradiation with a high radiation dose in cultured cells or organisms that had been pre-treated with a priming low radiation dose. The priming low radiation dose is usually called “adapting dose” or “conditioning dose” while the high radiation dose is called “challenge dose”. The induction of radioadaptive response was first reported by Olivieri *et al.* (1984) who showed that the frequency of chromatid aberrations were down to 50% less than expected after exposure to 1.5 Gy of x-rays. Many articles have demonstrated radioadaptive response in plant cells (Cortes *et al.* 1990), insects (Fritz-Niggli and Schaeppi-Buechi 1991), Chinese hamster V79 cells (Ikushima 1987, 1989a, 1989b), cultured human lymphocytes (Shadley and Wolff 1987; Wolff *et al.* 1988; Sankaranarayanan *et al.* 1989; Shadley and Wiencke 1989; Ghiassi-nejad *et al.* 2002), embryonic and HeLa cells (Ishii and Watanabe 1996), occupationally exposed persons (Barquinero *et al.* 1995; Gourabi and Mozdarani 1998), cultured animal lymphocytes (Wojcik and Tuschl 1990), and in vivo studies on laboratory animals (Bosi and Olivieri 1989; Cai and Liu 1990; Liu *et al.* 1992; Farooqi and Kesavan 1993). However, there are reports indicating lack of radioadaptive response in cultured human lymphocytes (Mortazavi *et al.* 2000; Mortazavi *et al.* 2003b; Joksic and Petrovic 2004). Furthermore, long-term follow up studies indicate that lack of radioadaptive response is not a temporary effect and, in contrast with the early reports of Olivieri and Bosi (1990), does not depend on transient physiological factors (Ikushima and Mortazavi 2000).

Although there is substantial evidence about the induction of adaptive response with low doses of ionizing radiation, doubt still persists concerning whether it is possible to induce such a response after exposure to adapting doses of non-ionizing radiations such as microwaves. Mobile phones, as wireless communication devices with drastically widespread use and increased popularity, use electromagnetic radiation in the microwave range. Although mobile phones and base stations (towers) produce the same type of radiation, phones play a much more significant role in human exposures than towers (Slesin 2000). It is reported that the output power of towers is two to three orders of magnitude greater than that of mobile phones. However, the phones are held just a few centimeters from some sensitive parts of the body such as the brain and the eyes, which causes a dramatic concern regarding the health effects of the microwaves. It is estimated that approximately 50-70% of the power output of a phone is absorbed by the user (Slesin 2000).

The aim of this study was to investigate the effects of irradiation of mice and rats with microwaves, as a priming or conditioning dose, on the

induction of adaptive response to a subsequent lethal dose (LD) of gamma rays.

MATERIALS & METHODS

Animals

In the 1st phase of the study, one hundred and ten male Balb/c mice (25-30 g) were randomly divided into 8 groups of 13-15 animals. In the 2nd phase, to investigate if exposure of another animal model to a challenge dose of a common LD 50/30 can induce the same survival adaptive response, thirty male adult Wistar rats (200-250 g) were randomly divided into 2 groups of 15 animals. The animals were kept in special cages with controlled temperature, humidity and lighting. All animal experiments were considered and approved by the Animal Experimentation Ethics Committee of Shiraz University of Medical Sciences prior to commencing work.

GSM mobile phone simulator

A GSM mobile phone simulator (designed and produced at the School of Electrical and Computer Engineering, Shiraz University in collaboration with the Center for Research in Radiation Sciences, Shiraz University of Medical Sciences) was used for microwave irradiation. The frequency was adjustable from 800 to 1000 MHz but in this experiment 900 MHz radiation (a wavelength of about 33.4 cm) that is similar to GSM mobile phone was used. Amplitude was digitally modulated with a 217Hz frequency and the duty cycle was 0.125 (the signal was switched on for 463 micro seconds periodically with a rate of 217 Hz. The power was adjustable from zero to over 3 Watts but we fixed it at 2 W during exposure. The signal bandwidth was 200kHz (similar to GSM mobile phone channels).

Adapting Dose (Microwave and Low Dose Rate Gamma Irradiation)

During the MW exposure, the animals were immobilized by placing their body through plastic restrainers. The distance between the antenna of the mobile simulator and animal's head was 10 cm. To expose the animals to LDRG radiation, a Cs-137 source with activity of 10 mCi was used. The dose rate for animals located at a distance of 10 cm was 2.6 mSv/h.

As summarized in Table 1, the 8 groups in the 1st phase of the study were as follows. The 1st group or controls received neither an adapting nor a challenge dose. The animals in the 2nd, 3rd and 4th groups were exposed only to adapting doses of microwave (MW) radiation, low dose rate gamma radiation (LDRG) or both (MW+LDRG), respectively. The animals in the 5th, 6th and 7th groups were exposed to adapting doses of MW, LDRG or both (MW+LDRG), respectively, and then irradiated with

TABLE 1. Survival rates in different groups of mice 6 and 12 days after exposure/sham exposure to the lethal dose (LD 50/6) of gamma radiation in the 1st phase of the study.

Groups	Treatment		Survival Rate	
	Adapting Dose	Challenge Dose (LD 50/6)	After 6 Days	After 12 Days
Group 1	-	-	95%	95%
Group 2	MW	-	95%	95%
Group 3	LDRG	-	85%	85%
Group 4	MW + LDRG	-	100%	100%
Group 5	MW	8.8 Gy	80%	60%
Group 6	LDRG	8.8 Gy	90%	90%
Group 7	MW + LDRG	8.8 Gy	90%	80%
Group 8	-	8.8 Gy	60%	10%

a challenge lethal dose of gamma radiation. LDRG irradiation was scheduled for 18 hours per day for 5 days (a subtotal dose of 46.8 mSv per day and a total dose of 234 mSv). Finally, animals in the 8th group received no adapting dose but were exposed to a challenge dose of gamma radiation. The animals in the appropriate groups were exposed to microwave radiation 6 hours per day for either 5 days (1st phase of the study), or six days (2nd phase) before exposure to the lethal dose of gamma radiation. In the 2nd phase, the animals in the 1st group received microwave exposure 6 hours per day for 4 days. The animals in the 2nd group received no microwave irradiation.

Challenge Lethal Dose (LD)

On day 6, exposure of the Balb/c mice to a previously reported (Hanson *et al.* 1987) lethal dose (LD 50/6) of 8.8 Gy of gamma rays was performed using a radiotherapy Cobalt-60 unit (35_35 cm² field, 48.54 cGy/min dose rate, 18.13 min irradiation time). In the 2nd phase, on day 5, all Wistar rats were whole-body irradiated with a previously reported (Lee *et al.* 1998) LD 50/30 of 8 Gy gamma radiation emitted by a radiotherapy Cobalt-60 source (35_35 cm² field, 54.68 cGy/min dose rate, 14.63 min irradiation time).

Assessment of Adaptive Response

Our basic measurement was the DR (Death Rate), calculated as (1 - survival fraction). In the next step, the expected DRs in animals that were pretreated with an adapting dose of MW, LDRG, or both (MW+LDRG) were calculated as:

$$DR_{\text{Expected}} = DR_{\text{RF}} \cdot LD_{\text{RG}} \cdot (\text{MW+LDRG}) + DR_{\text{LD50}} - DR_{\text{Controls}} \quad (1)$$

where, DR_{RF} , LD_{RG} , (MW+LDRG) is inverse survival in animals only exposed to adapting doses of MW, LDRG, or both (MW+LDRG), without subsequent irradiation with lethal dose (LD50). $DRLD50$ is inverse survival in animals only exposed to a challenge lethal dose without pre-irradiation with adapting doses of MW, LDRG, or both (MW+LDRG).

The coefficient of induced adaptive response, k , in each experiment was calculated as follows:

$$k = DR_{\text{Observed}} / DR_{\text{Expected}} \quad (2)$$

where, DR_{Observed} is inverse survival in animals not only exposed to adapting doses of MW, low dose rate gamma (LDRG), or both (MW+LDRG), but also subsequently irradiated with a lethal dose. A k value of less than one indicates a radioadaptive response. If $k = 1$, it means that there is no radioadaptive (a simple additivity) effect. Finally, if k is significantly greater than 1, it means that a synergistic effect¹ was induced.

Survival Study

After gamma irradiation, the animals were monitored either for 12 days (1st phase, animals that were exposed to LD 50/6) or 30 days (2nd phase, animals that were exposed to LD 50/30). During these two intervals, the general clinical state of the living animals as well as death events were controlled and recorded daily by an expert group.

Statistical Analysis

Kaplan-Meier's survival analysis was used for assessing the survival rate in each group. A dead animal was counted as 0, whereas alive animals were defined as 1. The difference among the survival rates of the groups was evaluated by the log-rank (Mantel-Cox) test. The statistical significance of the log-rank test was considered if $p < 0.05$ according to chi-square test.

RESULTS

In the 1st phase of the study, as shown in Table 1, six days after exposing the mice to the LD 50/6, the survival rate in the 1st group (controls) that received neither adapting nor challenge dose was 95%. This rate in the animals of the 2nd, 3rd and 4th groups that were exposed only to adapting doses of microwave radiation, LDRG or both (MW+LDRG), was 95%, 85% and 100%, respectively. Survival rates in the animals of the 5th, 6th and 7th groups that were first exposed to conditioning doses of

¹Synergism means the observed effect of exposure to a combination of substances is greater than the sum of the effects of each substance administered individually (e.g. $2+2 = 5$).

microwave radiation, LDRG or both (MW+LDRG) and then irradiated with an LD 50/6 dose of gamma radiation were 80%, 90% and 90%, respectively. Finally, the survival rate in the animals of the 8th group that received no adapting dose but were exposed to the LD 50/6 dose was 60%.

As shown in Table 2, twelve days after exposing/sham exposing the mice to the LD 50/6 dose, the survival rate in the 1st group (controls) was still 95%. This rate in the animals of the 2nd, 3rd and 4th groups was 95%, 85% and 100%, respectively. Survival rate in the animals of the 5th, 6th and 7th groups was 60%, 90% and 80%, respectively. Finally, the survival rate in animals of the 8th group was only 10%. When overall animal survival data were evaluated using the Kaplan-Meier test, a statistically significant difference among survival rates of the mice in different groups was observed. Furthermore, statistically significant differences between the survival rate of the mice only exposed to lethal dose of gamma radiation before irradiation with a lethal dose of gamma radiation with those of the animals pre-exposed to either microwave ($p=0.02$), low dose rate gamma ($p=0.001$) or both of these physical adapting doses ($p=0.003$) were observed. Kaplan-Meier survival plots of the mice pre-exposed/sham-exposed to microwave, low dose rate gamma or both of these adapting doses before irradiation with a lethal dose (LD) of gamma radiation is shown in Figure 1.

In the 2nd phase of the study, 30 days after irradiation of the rats, the survival fractions for the control group (no pretreatment before LD), was 53.3%, while there was no death event (survival rate of 100%) in the animals receiving microwave radiation before the LD 50/30 (Table 3). Alteration of the survival rates of the rats pre-exposed to microwave (MW) before irradiation with a lethal dose (LD) of gamma radiation compared to those only exposed to lethal dose (No microwave pre-irradiation) is shown in Figure 2.

TABLE 2. Observed and expected survival rates and the coefficient of the induced adaptive response in different groups of mice on day 12 in the 1st phase of the study.

Groups	Treatment		Death Rate (1-Survival Rate)		Coefficient of Adaptive Response (Observed/Expected)
	Adapting Dose (AD)	Challenge Dose (CD)	Observed	Expected (AD+CD-cont)	
Group 1	-	-	0.05	-	
Group 2	MW	-	0.05	-	
Group 3	LDRG	-	0.15	-	
Group 4	MW + LDRG	-	0	-	
Group 5	MW	LD 50/6	0.20	0.40	0.50
Group 6	LDRG	LD 50/6	0.10	0.50	0.20
Group 7	MW + LDRG	LD 50/6	0.10	0.35	0.29
Group 8	-	LD 50/6	0.40	-	

Microwave Radiation and Survival Adaptive Response

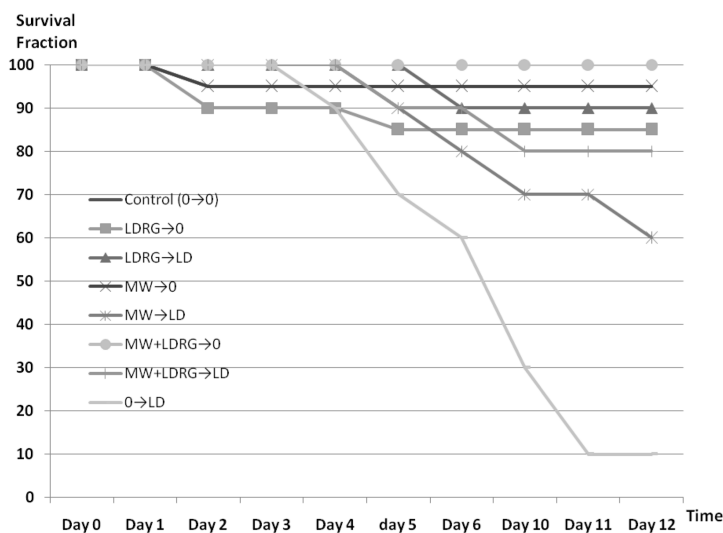


FIGURE 1. Kaplan-Meier survival curves of the mice pre-exposed/sham-exposed to microwave (MW), low dose rate gamma (LDRG) or both (MW+LDRG) before irradiation with a lethal dose (LD) of gamma radiation.

TABLE 3. Survival rates in different groups of rats 10, 20 and 30 days after exposure/sham exposure to the lethal dose (LD 50/30) of gamma radiation in the 2nd phase of the study.

Groups	Treatment		Survival Rate After:		
	Adapting Dose	Challenge Dose	10 Days	20 Days	30 Days
Group 1	MW 12:3	LD 50/30	100%	100%	100%
Group 2	-	LD 50/30	100%	86.6%	53.3%

MW 12:3 = Microwave irradiation for 3 hours every 12 hours (6 h/day)

DISCUSSION

An earlier exposure of the cells or tissues to a small dose of ionizing radiation usually increases their resistance to toxicity caused by a high dose radiation; a phenomenon that is usually called adaptive or radioadaptive response. Results obtained in both phases of this study clearly indicate the possibility of the induction of survival adaptive response as increased resistance to lethal doses of gamma rays by pre-exposing the animals to microwave radiation.

In the 1st phase of the study, 6 days after exposing the mice to the LD 50/6, the survival rate in the controls (no pretreatment, only LD) was 60% (a reasonably good level of agreement between our results with the previously published values for LD 50/6), while the survival rates in the animals pretreated with LDRG, microwave radiation and both LDRG and microwave radiation, were 90%, 80% and 90%, respectively. In the 2nd

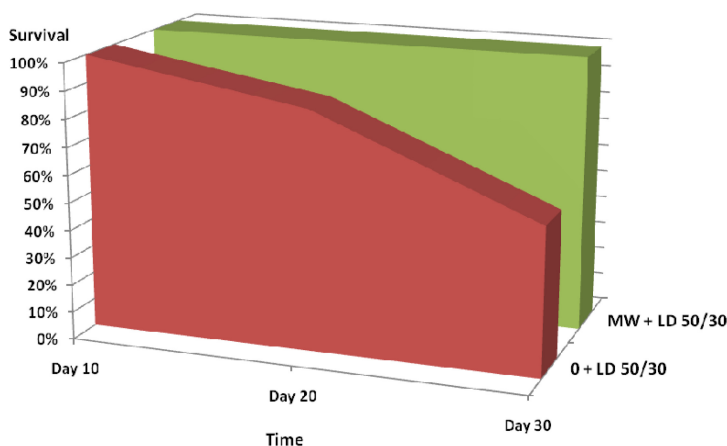


FIGURE 2. Alteration of the survival rates of the rats pre-exposed to microwave (MW) before irradiation with a lethal dose (LD) of gamma radiation compared to those only exposed to lethal dose (No microwave pre-irradiation).

phase of the study, survival fractions, 30 days after irradiation of the animals with LD 50/30, was 100% for the animals pre-treated with microwave radiation, while it was 53.3% for the animals that had no pre-treatment before LD 50/30 (control group).

These findings are in line with the very limited recently published reports that indicated the possibility of the induction of adaptive response after pre-treatment with microwave radiation (Sannino *et al.* 2009; Cao *et al.* 2011; Sannino *et al.* 2011; Jiang *et al.* 2012; Zeni *et al.* 2012). Sannino *et al.* (2009) have previously reported that pre-exposure of peripheral blood lymphocytes collected from human volunteers to non-ionizing RF radiation (900 MHz, at a peak specific absorption rate of 10 W/kg for 20 h) increases their resistance to a challenge dose of mitomycin C (100 ng/ml at 48 h). Later, they confirmed their previous results and showed that the timing of adapting dose exposure of radiofrequency plays an important role in the process of adaptive response induction (Sannino *et al.* 2011). On the other hand, Chinese researchers have recently shown that pre-exposure of mice to non-ionizing 900 MHz RF induced adaptive response and thus reduced the hematopoietic tissue damage from a subsequent challenge dose of ionizing radiation (Cao *et al.* 2011).

Our results are in line with those reported recently by Zeni *et al.* (2012) who showed that when lymphocytes were pre-exposed to RF at 0.3W/kg SAR and then treated with mitomycin C, these cells showed a significant reduction in the frequency of micronuclei, compared with the cells treated with MMC alone. Jiang *et al.* (2012) also recently used a relatively similar method as we did previously (using the gamma radiation as the challenge dose) and showed that mice pre-exposed to RF for 3, 5, 7

and 14 days showed progressively decreased damage and were significantly different from those exposed to gamma-radiation alone. In this light, in spite of the fact that there were some basic differences between our study and the above-mentioned investigation, both studies confirm the potential of RF radiation for induction of adaptive response. The cardinal differences between our study and that conducted by Sannino *et al.* (2009) include the type of the challenge dose ; i.e. lethal dose of gamma radiation in our study versus chemical agent of mitomycin C in the study by Sannino *et al.* (2009) and the endpoint; i.e. survival rate, 30 days after LD 50/30 in our study versus induction of micronuclei in the study by Sannino *et al.* (2009).

Some recent studies showed that radiofrequency radiation causes oxidative injury in different tissues mediated by lipid peroxidation, increased level of nitric oxide (NO) and suppression of antioxidant defense mechanism (Ozgun *et al.* 2010; Esmekaya *et al.* 2011). Feinendegen *et al.* in their previous studies (1996, 1999), proposed that adaptive response could be induced by reactive oxygen species (ROS). Increased levels of ROS or NO have been usually observed in adapted cells (Tapio and Jacob 2007). The ROS refers to a group of molecules including peroxides and free radicals which are derived from oxygen and are highly reactive toward biomolecules (Maynard *et al.* 2009). ROS react with critical biomolecules such as DNA and induce oxidative stress (imbalance of prooxidants versus antioxidants) and damage in these macromolecules, multiple localized lesions such as base damage, single strand breaks (SSBs) and double strand breaks (DSBs), DNA-DNA cross links and DNA-protein cross links (Goldberg and Lehnert 2002; Marnett *et al.* 2003; Cejas *et al.* 2004; Murray *et al.* 2005). As indicated by some investigators (Bose Girigoswami and Ghosh 2005; Yan *et al.* 2006; Dimova *et al.* 2008), we believe that induction of adaptive response by pre-exposure to ionizing radiation needs a minimum level of damage that triggers this phenomenon and increases the resistance of living organisms (in vivo) or cells (in vitro) to higher levels of the same or of other sources of stress.

Our findings also confirmed that pre-irradiation of the animals with dose rates of gamma rays as low as 2.6 mSv/h (46.8 mSv/day, total dose of 234 mSv) as priming or conditioning dose significantly increases the radioresistance of animals. This finding is in line with the results reported by Elmore *et al.* (2008) who showed that a dose of 100 mGy at dose rates of 1-4 mGy/day was able to induce an adaptive response, while at dose rates below about 1 mGy/day the adaptive response was lost. This suggests that the lower dose threshold for adaptation depends on the presence of a minimum number of lesions per unit time.

It should be noted that in the case of ionizing radiation, there are reports indicating lack of radioadaptive response in cultured lymphocytes

of some individuals (Bosi and Olivieri 1989; Olivieri and Bosi 1990; Hain *et al.*,1992). Specially when long-term follow up studies of Mortazavi *et al.*, which indicate that lack of radioadaptive response is not a temporary effect and does not depend on transient physiological factors (Ikushima and Mortazavi 2000; Mortazavi *et al.* 2000, 2003a), are taken into account, further studies are needed to clarify if the same pattern of variability exists in non-ionizing, radiation-induced adaptive response. Extending these data to humans would suggest that pre-exposure to electromagnetic radiation in the microwave region may decrease the therapeutic outcome of radiation therapy. On the other hand, this phenomenon can save the lives of humans who are exposed to life threatening doses of ionizing radiation, eg, astronauts who are selected for deep space missions.

CONCLUSIONS

The results obtained in this study clearly show that exposure of laboratory animals to microwave irradiation can induce a survival adaptive response as increased survival fraction at a specific time after exposure to a lethal dose of radiation. Although this phenomenon may have a wide variety of applications in radiation protection or radiation biology, the radioresistance induced by microwave radiations emitted by common sources of microwave radiation such as mobile phones may interfere with the outcome of any subsequent therapeutic application of radiation in radiotherapy. In this light, further studies are needed to clarify the extent of this effect and its persistence.

CONFLICT OF INTEREST

None

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REFERENCES

- Barquinero JF, Barrios L, Caballin MR, Miro R, Ribas M, Subias A, Egozcue J. 1995. Occupational exposure to radiation induces an adaptive response in human lymphocytes. *Int J Radiat Biol* 67:187-191.
- Bose Girigoswami K, Ghosh R. 2005. Response to gamma-irradiation in V79 cells conditioned by repeated treatment with low doses of hydrogen peroxide. *Radiat Environ Biophys* 44:131-137.
- Bosi A, Olivieri G. 1989. Variability of the adaptive response to ionizing radiations in humans. *Mutat Res* 211:13-17.
- Cai L, Liu SZ. 1990. Induction of cytogenetic adaptive response of somatic and germ cells in vivo and in vitro by low-dose X-irradiation. *Int J Radiat Biol* 58:187-194.

- Cao Y, Xu Q, Jin ZD, Zhou Z, Nie JH, Tong J. 2011. Induction of adaptive response: Pre-exposure of mice to 900 MHz radiofrequency fields reduces hematopoietic damage caused by subsequent exposure to ionising radiation. *Int J Radiat Biol* 87:720-728.
- Cejas P, Casado E, Belda-Iniesta C, De Castro J, Espinosa E, Redondo A, Sereno M, Garcia-Cabezas MA, Vara JA, Dominguez-Caceres A, Perona R, Gonzalez-Baron M. 2004. Implications of oxidative stress and cell membrane lipid peroxidation in human cancer (Spain). *Cancer Causes Control* 15:707-719.
- Cortes F, Dominguez I, Mateos S, Pinero J, Mateos JC. 1990. Evidence for an adaptive response to radiation damage in plant cells conditioned with X-rays or incorporated tritium. *Int J Radiat Biol* 57:537-541.
- Dimova EG, Bryant PE, Chankova SG. 2008. Adaptive response: some underlying mechanisms and open questions. *Genet Mol Biol* 31:396-408.
- Elmore E, Lao XY, Kapadia R, Giedzinski E, Limoli C, Redpath JL. 2008. Low doses of very low-dose-rate low-LET radiation suppress radiation-induced neoplastic transformation in vitro and induce an adaptive response. *Radiat Res* 169:311-318.
- Esmekaya MA, Ozer C, Seyhan N. 2011. 900 MHz pulse-modulated radiofrequency radiation induces oxidative stress on heart, lung, testis and liver tissues. *Gen Physiol Biophys* 30:84-89.
- Farooqi Z, Kesavan PC. 1993. Low-dose radiation-induced adaptive response in bone marrow cells of mice. *Mutat Res* 302:83-89.
- Feinendegen LE, Bond VP, Sondhaus CA, Altman KI. 1999. Cellular signal adaptation with damage control at low doses versus the predominance of DNA damage at high doses. *C R Acad Sci III* 322:245-251.
- Feinendegen LE, Bond VP, Sondhaus CA, Muehlensiepen H. 1996. Radiation effects induced by low doses in complex tissue and their relation to cellular adaptive responses. *Mutat Res* 358:199-205.
- Fritz-Niggli H, Schaeppi-Buechi C. 1991. Adaptive response to dominant lethality of mature (class A) and immature (class B) oocytes of *D. melanogaster* to low doses of ionizing radiation: effects in repair-proficient (yw) and repair-deficient strains (mei 41D5 and mus 302D1). *Int J Radiat Biol* 59:175-184.
- Ghiassi-nejad M, Mortazavi SMJ, Cameron JR, Niroomand-rad A, Karam PA. 2002. Very high background radiation areas of Ramsar, Iran: Preliminary biological studies. *Health Physics* 82:87-93.
- Goldberg Z, Lehnert BE. 2002. Radiation-induced effects in unirradiated cells: a review and implications in cancer. *Int J Oncol* 21:337-349.
- Gourabi H, Mozdarani H. 1998. A cytokinesis-blocked micronucleus study of the radioadaptive response of lymphocytes of individuals occupationally exposed to chronic doses of radiation. *Mutagenesis* 13:475-480.
- Hain J, Jaussi R, Burkart W. 1992. Lack of adaptive response to low doses of ionizing radiation in human lymphocytes from five different donors. *Mutat Res* 283:137-144.
- Hanson WR, Fry RJ, Sallase AR, Frischer H, Ahmad T, Ainsworth EJ. 1987. Comparison of intestine and bone marrow radiosensitivity of the B6ALB/c and the C57BL/6 mouse strains and their B6CF1 offspring. *Radiat Res* 110:340-352.
- Ikushima T. 1987. Chromosomal responses to ionizing radiation reminiscent of an adaptive response in cultured Chinese hamster cells. *Mutat Res* 180:215-221.
- Ikushima T. 1989a. Radio-adaptive response: characterization of a cytogenetic repair induced by low-level ionizing radiation in cultured Chinese hamster cells. *Mutat Res* 227:241-246.
- Ikushima T, editor. 1989b. Radioadaptive response: a novel chromosomal response in Chinese hamster cells in vitro. Berlin: Springer-Verlag.
- Ikushima T, Mortazavi SMJ. 2000. Radioadaptive response: its variability in cultured human lymphocytes. In: Yamada T, Mothersil C, Michael BD, Potten CS, editors. *Biological Effects of Low Dose Radiation*. Amsterdam: Elsevier. p 81-86. .
- Ishii K, Watanabe M. 1996. Participation of gap-junctional cell communication on the adaptive response in human cells induced by low dose of X-rays. *Int J Radiat Biol* 69:291-299.
- Jiang B, Nie J, Zhou Z, Zhang J, Tong J, Cao Y. 2012. Adaptive response in mice exposed to 900 MHz radiofrequency fields: primary DNA damage. *PLoS One* 7:e32040.
- Joksic G, Petrovic S. 2004. Lack of adaptive response of human lymphocytes exposed in vivo to low doses of ionizing radiation. *J Environ Pathol Toxicol Oncol* 23:195-206.
- Lee YK, Chang HH, Kim WR, Kim JK, Yoon YD. 1998. Effects of gamma-radiation on ovarian follicles. *Arh Hig Rada Toksikol* 49:147-153.

- Liu SZ, Cai L, Sun SQ. 1992. Induction of a cytogenetic adaptive response by exposure of rabbits to very low dose-rate gamma-radiation. *Int J Radiat Biol* 62:187-190.
- Marnett LJ, Riggins JN, West JD. 2003. Endogenous generation of reactive oxidants and electrophiles and their reactions with DNA and protein. *J Clin Invest* 111:583-593.
- Maynard S, Schurman SH, Harboe C, de Souza-Pinto NC, Bohr VA. 2009. Base excision repair of oxidative DNA damage and association with cancer and aging. *Carcinogenesis* 30:2-10.
- Mortazavi S, Ikushima T, Mozdarani H. 2003a. Variability of chromosomal radioadaptive response in human lymphocytes. *Iran J Radiat Res* 1:55-61.
- Mortazavi SMJ, Cameron JR, Niroomand-rad A. 2003b. Adaptive response studies may help choose astronauts for long-term space travel. *Adv Space Res* 31:1543-1551.
- Mortazavi SMJ, Ikuhima T, Mozdarani H, Sharafi AA, Ishi Y. 2000. Is low-level pre-irradiation of human lymphocytes an absolutely beneficial phenomenon. A report on the extra-ordinary synergism. *Kowsar Medical Journal* 5:235-240.
- Murray D, Allalunis-Turner M, Weinfeld M. 2005. VIIIth International Workshop on Radiation Damage to DNA. *Int J Radiat Biol* 81:327-337.
- Olivieri G, Bodycote J, Wolff S. 1984. Adaptive response of human lymphocytes to low concentrations of radioactive thymidine. *Science* 223:594-597.
- Olivieri G, Bosi A, editors. 1990. Possible causes of variability of the adaptive response in human lymphocytes.
- Ozgur E, Guler G, Seyhan N. 2010. Mobile phone radiation-induced free radical damage in the liver is inhibited by the antioxidants N-acetyl cysteine and epigallocatechin-gallate. *Int J Radiat Biol* 86:935-945.
- Sankaranarayanan K, von Duyn A, Loos MJ, Natarajan AT. 1989. Adaptive response of human lymphocytes to low-level radiation from radioisotopes or X-rays. *Mutat Res* 211:7-12.
- Sannino A, Sarti M, Reddy SB, Prihoda TJ, Vijayalaxmi, Scarfi MR. 2009. Induction of adaptive response in human blood lymphocytes exposed to radiofrequency radiation. *Radiat Res* 171:735-742.
- Sannino A, Zeni O, Sarti M, Romeo S, Reddy SB, Belisario MA, Prihoda TJ, Vijayalaxmi, Scarfi MR. 2011. Induction of adaptive response in human blood lymphocytes exposed to 900 MHz radiofrequency fields: Influence of cell cycle. *Int J Radiat Biol* 87: 993-9.
- Shadley JD, Wiencke JK. 1989. Induction of the adaptive response by X-rays is dependent on radiation intensity. *Int J Radiat Biol* 56:107-118.
- Shadley JD, Wolff S. 1987. Very low doses of X-rays can cause human lymphocytes to become less susceptible to ionizing radiation. *Mutagenesis* 2:95-96.
- Slesin L. 2000. Public Concerns over Microwave Radiation in the U.S.: Comparing the Perceived Health Risks of Phones and Towers. In: International Conference on Cell Tower Siting Linking Science & Public Health. Salzburg. p 173-176.
- Tapio S, Jacob V. 2007. Radioadaptive response revisited. *Radiat Environ Biophys* 46:1-12.
- Wojcik A, Tuschl H. 1990. Indications of an adaptive response in C57BL mice pre-exposed in vivo to low doses of ionizing radiation. *Mutat Res* 243:67-73.
- Wolff S, Afzal V, Wiencke JK, Olivieri G, Michaeli A. 1988. Human lymphocytes exposed to low doses of ionizing radiations become refractory to high doses of radiation as well as to chemical mutagens that induce double-strand breaks in DNA. *Int J Radiat Biol Relat Stud Phys Chem Med* 53:39-47.
- Yan G, Hua Z, Du G, Chen J. 2006. Adaptive response of *Bacillus* sp. F26 to hydrogen peroxide and menadione. *Curr Microbiol* 52:238-242.
- Zeni O, Sannino A, Romeo S, Massa R, Sarti M, Reddy AB, Prihoda TJ, Vijayalaxmi, Scarfi MR. 2012. Induction of an adaptive response in human blood lymphocytes exposed to radiofrequency fields: Influence of the universal mobile telecommunication system (UMTS) signal and the specific absorption rate. *Mutat Res*. [Epub ahead of print]